



**Annual Research & Review in Biology**  
4(9): 1387-1396, 2014

SCIENCEDOMAIN *international*  
[www.sciencedomain.org](http://www.sciencedomain.org)



---

# Mesenchymal Stem Cell-based Cancer Gene Therapy: Application and Unresolved Problems

Sinh Truong Nguyen<sup>1</sup>, Viet Quoc Pham<sup>1</sup>, Ngoc Kim Phan<sup>1</sup>  
and Phuc Van Pham<sup>1\*</sup>

<sup>1</sup>Laboratory of Stem Cell Research and Application, University of Science, Vietnam National University, Ho Chi Minh city, Vietnam.

## Authors' contributions

*This work was carried out in collaboration between all authors. Authors STN and VQP wrote the parts of 1, 2 and 3. Authors NKP and PVP wrote the parts of 4, 5 and edited manuscript. All authors read and approved the final manuscript.*

Review Article

Received 1<sup>st</sup> October 2013  
Accepted 7<sup>th</sup> January 2014  
Published 13<sup>th</sup> January 2014

---

## ABSTRACT

There are many barriers that have hindered the progress of clinical application of gene therapy as a potential method for treating cancer. These limitations are caused by transduction/delivery failure rates with viral vector systems and were due to the resultant non-specific targeting as well as the triggering of immune system stimulation. Recently, mesenchymal stem cells (MSCs) were found to be advantageous in improving tumor tropism as well as in providing an immune-privilege. Therefore, the combination of viral vector systems and stem cells was suggested as an attractive solution to promote targeted-delivery of anticancer therapeutics to tumor sites. In this regard, many published studies have shown that MSCs could be useful as a potential vehicle for delivering anticancer agents to tumors. In this review, the disadvantages of viral vector systems for gene therapy were analyzed, and an updated account on the role of MSCs in cancer gene therapy was presented. In addition, major safety and therapeutic unresolved problems facing this approach in clinical application were also addressed.

**Keywords:** *Mesenchymal stem cells; gene therapy; tumor tropism; immune-privilege; cancer.*

---

\*Corresponding author: Email: [pvphuc@hcmuns.edu.vn](mailto:pvphuc@hcmuns.edu.vn);

## 1. INTRODUCTION

Gene therapy can be classified into two types *ex vivo* (where the cell is manipulated outside the body and transplanted back into the patient's body again) and *in vivo* (where the desired gene is transferred inside the patient's body).

Most of the current *in vivo* studies of cancer gene therapy applications state that the desired genes must enter the cancer cell and function directly or indirectly inside the patient. Transformation of genes requires a carrier vehicle called the *Vector*. Gene delivery method by vectors can be classified into two: *viral* and *non-viral vectors*. *Viral vectors* such as *Herpes Simplex Virus* have been used to successfully treat animal disease models [1-5], cancer [6-10] and chronic pain [11-13]. In the case of *adenoviral s vectors*, the first clinical trials of this virus therapy for cystic fibrosis received no evidence for clinical efficacy, but more clinical studies are committed with adenovirus vectors for the treatment of cancer, some examples include clinical trials on glioblastoma [14], breast cancer [15], gynecologic cancer [16], colorectal carcinoma [10], hepatocellular carcinoma [17] and myocardial angiogenesis [18]. However, *viral vectors* used in targeting cancer cell in the patient's body may have resulted in nonspecific genomic rearrangements in normal cells and had an impact on cell physiology at the integration site. The limitation of this vector includes the stimulation of host immune system.

In the case of *ex vivo* gene therapy, therapeutic cells are modified outside the patient's body to formulate the desired genes carriers, which are then transfected into patients. The types of therapeutic cell that were used in gene therapy treatment include hematopoietic stem cells, neural stem cells, and mesenchymal stem cells. These kinds of stem cells have been demonstrated to possess the ability to suppress host immune system with the resulting efficiency of carrying therapeutic gene to the desired site within the patient's body. Therefore, a combination of gene and stem cell therapy would be a better choice in overcoming the barriers of gene therapy for cancer treatment.

## 2. SHORTCOMING OF GENE THERAPY

There are two major obstacles that are known to limit the clinical efficacy of gene therapy applications: lack of vector- tumor tropism and the triggering of an immune response.

The human immune system has evolved to fight off foreign antigens that are introduced into the body. This mechanism helps to maintain genetic fidelity and stability throughout the human evolution. Therefore, innate immunity and antigen-specific adaptive immune responses against vector-derived antigens may reduce the efficacy and stability of an *in vivo* gene transfer. In addition, some vectors are derived from human viruses leading memory immune responses against vector antigens. Moreover, antibody and T-cell responses may be directed against therapeutic gene products that may differ from endogenous proteins [19-21]. Therefore, immune stimulation targeting viral vectors is always the main obstacle for a successful gene therapy.

The majority of gene therapy clinical trials were dependent on viral vector systems to transduce exogenous genes into target cells. The choice of viral vectors may be due to their efficient transducing powers; however, these viral vectors will transduce genes in a non-specific manner. These viral vectors will transduce therapeutic genes non- differentially into both target and non-target cells, which will induce toxicity for normal body cells. For

example, transduction of adenovirus-mediated gene therapy was based on the high-affinity binding of the knob domain of the virus fiber protein in Coxsackie-adenovirus receptor (CAR) to the membrane of the target cell [22,23], however, when the cancer cell down-regulate CAR on their membrane, the specific-transducing ability of the virus would be decreased [23]. In addition, CAR is localized to epithelial tight junctions in vivo with its responsibility in the regulation of epithelial permeability and tissue homeostasis [24], therefore, it will be non-feasible to use CAR as a specific marker to safely and efficiently deliver therapeutic genes into target cells. It was evident that receptor specificity for the efficient transduction of these therapeutic genes is lacking and constitutes a barrier for the widespread safety in the application of adenovirus vectors; hence, the binding between a virus ligand and its cellular receptor is a crucial and important step for a viral infection. Studies aimed at overcoming the safety problems caused by the failure of natural viral vector tropism would need more time to show better and safe outcomes. These unresolved application barriers have obstructed the use of current vectors as specific and efficient targeting tools/ models in cancer therapy. The major aim for future gene therapy studies is to incorporate stem cells as an attractive delivery option targeted at providing potential protection for viral vectors from immune surveillance in support of any targeted delivery of genes or therapeutic proteins to tumor sites.

### **3. MESENCHYMAL STEM CELL-BASED CANCER THERAPY**

Mesenchymal stem cells are stem cells that have shown great interest due to their apparent ability to home to injury sites following their systemic delivery. It is evident that MSCs possess an inherent ability to both self-renew and differentiate into multiple lineages including osteoblasts, chondrocytes and adipocytes. These cells can easily be isolated from many sources including the stromal compartments of bone marrow, adipose tissue, trabecular bone and skeletal muscles.

Mesenchymal stem cells have the ability to migrate to a variety of organs and tissues when injected, however, localized pathogen-induced inflammatory site will elicit enhanced MSCs homing for general remodeling and repair. Recent reports have shown that MSCs preferentially migrate to sites of injury in an animal model of ischemia [25], central nervous system [26,27], lung [28], liver [29], musculoskeletal tissue[30], ischemic brain [31], kidney [32] and glioma [33].

The tumor homing ability of MSCs seems to be based on chemo-attraction. In fact, MSCs are known to express chemokine and chemokine receptors that can be up-regulated by cytokines. It has been demonstrated that MSCs express a wide range of mRNA encoding cytokines, chemokines and their receptors. Tumor products such as CXCL12/stromal cell derived factor alpha, CX3CL1/fractalkine, CXCL10/interferon-gamma inducible protein play a significant role in their migration [34]. Pro-inflammatory factors including TNF-alpha and IFN-gamma are key in regulating the expression of chemokines and their receptors in MSCs [34, 35]. Interleukin IL-1 $\beta$ , a representative inflammatory factor, has been demonstrated to induce the secretion of trophic factors and adhesion to extracellular matrix components such as collagen and laminin in MSCs [36] will support their adhesion and migration processes. Therefore, expressing chemokines and their receptors in MSCs is considered the key mediators that provide linkage to tumor tropism. Recently, soluble factors produced by macrophages including IL-8, CCL-2, CCL-5 were shown to impact the mobility of MSCs in response to inflammation [37]. Fibronectin-rich matrices at vascular re-modelling sites may also play a key role in their tumor tropism. In fact, adhesion of MSCs with fibronectin has been demonstrated to activate platelet-derived growth factor receptor (PDGFR- $\beta$ )

phosphorylation in  $\alpha 5\beta 1$ -integrin dependent manner, resulting in actin reorganization and MSC migration [38].

Another functional advantage of MSCs is their capacity to elicit self-protective immunosuppressive effects upon their injection into the body. This property is due to the low expression of class II Major Histocompatibility Complex and co-stimulatory molecules on their surface throughout the different pathways of the immune response. This process is achieved by means of direct cell-to-cell interactions and soluble factors secretion, where MSCs can effectively target the main immune cell subsets to inhibit the proliferation of T-cells, B-cells, natural killer cells (NK) and dendritic cell (DC) [39,40]. It is well known that MSCs had the ability to reduce the inflammatory response of peripheral blood mononuclear cells through down-regulating the biological function of T-cell helper 17 (Th17) and inhibiting the expression of IL-7, IL-6 and TNF-alpha [41]. These Th17 cells are known to synthesize signature cytokine IL-17A to mediate localized tissue inflammation. It has been shown that MSCs inhibit Th17 differentiation from naive and memory T-cells as well as Th17 cells from the site of inflammation. The MSCs-derived prostaglandin E2 has been shown to mediate a suppressive effect on Th17 via the EP4 receptor [42]. Interestingly, this inhibitory effect of MSCs was found in inflammatory but not in healthy environments [41]. This pattern of events will support the fact that inflammatory mediators may have cause the immune regulatory property of MSCs. In fact, inflammatory cytokine-induced immunosuppression property of MSCs is manifested through adhesion molecules; MSCs were found to up-regulate the adhesive capacity of T-cells due to overexpression of ICAM-1 and VCAM-1 when co-cultured with T-cells. Hence, the more adhesion molecules that MSCs express, the more immune-suppression properties they possess [43]. Recent studies have shown that galectin-1, a protein detected intracellularly and on the cell surface of MSCs, plays an important role in inhibiting immune effector cells. Blocking of galectin-1 has led to loss of immune-regulatory ability of MSCs [44]. Moreover, MSCs-derived galectin-1 effectively regulated the release of cytokines that have influenced autoimmunity such as TNF $\alpha$ , IFN $\gamma$ , IL-2, and IL-10. This role provides an interesting novel function for adhesion molecules in MSCs besides their original adhesive functions. In addition, MSCs that were primed with IFN- $\gamma$  have induced cancer apoptosis through the tumor necrosis factor apoptosis-inducing ligand (TRAIL) [45]. This situation will provide a potential for enhancing the immune suppression ability of engineered MSCs that were primed with inflammatory agent such as IFN- $\gamma$ .

It is of utmost interest to emphasize that MSCs have the ability to carry their immune-modulating factors throughout the immune system while reaching for tumor sites. It has been shown that MSCs expressing LIGH; a member of TNF family as well as an immune stimulator, will eventually home to the tumor site and reverse the immune-suppressive microenvironment with the activation of CD4 and CD8 T cell response [46]. It is through the exploitation of these advantages, that MSCs were used to target tumors by carrying genes encoding IL-12 [47-49], IL-21 [50,51] in ovarian and retinal cancers. Another interleukin being suggested as an effective anti-tumor agent, IL-24 has been carried effectively by MSCs to target lung cancer sites. The interleukin transduced-MSCs were found to induce apoptosis and cell cycle arrest in cancer cells. The angiogenesis inhibitor PEDF-transduced MSCs were used to inhibit angiogenesis in gliomas [52] as well as Lewis lung cancer [53].

The tumor tropism and immune regulatory role of MSCs provides the perfect vehicle for carrying therapeutic gene to tumor sites without exposing them to the immune system. The transport of suicide genes has been associated with stem cells and has served as a potential for targeted cancer therapy. In this therapy, with the advantages of MSCs, suicide genes were carried to tumor sites by MSCs and the treatment activated pro-drugs into drugs that

killed the tumor cells. Recently, many studies have experimented with MSCs transduced with suicide genes from herpes simplex virus-thymidine kinase [54-56], and cytosine deaminase [57-62] to activate the pro-drug ganciclovir at the tumor site in attempting to reduce tumor size; for example, suicide gene transduced-MSCs associated with adjuvants such as valproic acid have enhanced their killing effects and has induced significantly tumor inhibition [63]. Recent studies that have experimented with MSCs in the delivery of therapeutic gene to treat multiple diseases including cancer by incorporation with lentivirus-mediated [56,64-70], retrovirus-mediated [71-74], plasmid mediated [75,76] and adenovirus mediated transduction [48,77,78]. Almost all the results in animal trials were found to prolong animal survival, reduce complications, and/or lower tumor volume.

#### **4. FUTURE OF STEM CELL-BASED CANCER GENE THERAPY**

In contrast to the above exciting news, recent studies have shown that MSCs may enhance the risk of tumor progression. The data in these studies showed that IL1 secreted from cancer cell promoted prostaglandin E of MSCs resulting in the secretion of proteins that have played a role in the inhibition of Th17. These interactions have led to the activation of beta-catenin signaling in cancer cell, resulting in the formation of a cancer stem cell niche via epithelial-mesenchymal transition [79,80]. In addition, MSCs may have provided a source of tumor-supporting microenvironment for osteosarcoma cells enhancing their ability for settlement and colonization that have resulted in tumor growth and metastasis and as well, served as a negative outcome in a rat model [81]. The functional interaction of cytokines and their receptors with extracellular matrix-receptors such as ICAM and VCAM have been suggested to play a positive role in tumor enhancement [81]. Moreover, galactin-1, the molecules expressed by MSCs and provides a role in the immune modulation property of MSCs, has been shown to be a critical factor for tumor promotion relating to the neo-vascularization process [82].

Furthermore, the metastasis of neuroblastoma is arguably caused by MSCs via the secretion of SDF-1 signaling [83] and the role of MSCs in enhancing the initiation of lung cancer also was also discussed [84,85]. The presence of MSCs in primary breast cancer tissues was confirmed and they were suggested to provide a favorable microenvironment for tumor cell growth [86]. Nonetheless, the immune-suppressive property of MSCs was demonstrated to favor tumor growth in allogeneic animal models [87]. These findings confirm that MSCs play a critical role in promoting cancer progression and therefore, the strategies of cancer gene therapy based on MSCs must raise a series of the question about the safety and counter productivity of the use of MSCs in cancer therapy.

#### **5. CONCLUSION**

The studies presented in this review have shown that stem cell-based gene therapy has a very promising potential for targeted cancer therapy. These cell-based applications have implicated interesting properties to overcome problems that were encountered with the use of conventional gene therapy. However, the above listed contrasting views and findings that relate to the use of MSC-based models of gene delivery in cancer therapy in connection with the stated problems in their application need to be underscored and thoroughly researched in the near future. These endeavors should attempt to provide extensive efforts to understand the underlying phenomenon and present research solutions in regard to the safety of biological applications of stem cell-based cancer gene therapy. This can be

achieved through in-depth preclinical studies that would maximize potential benefits while reducing the safety risks of these bio therapeutic applications in cancer.

## **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

## **REFERENCES**

1. Nygardas M, et al. Treatment of experimental autoimmune encephalomyelitis in SJL/J mice with a replicative HSV-1 vector expressing interleukin-5. *Gene Ther.* 2011;18(7):646-55.
2. Corona JC, et al. Hexokinase II gene transfer protects against neurodegeneration in the rotenone and MPTP mouse models of Parkinson's disease. *J Neurosci Res.* 2010;88(9):1943-50.
3. Brown WD, Bearer EL and Donahue JE. Chronic active herpes simplex type 2 encephalitis in an asymptomatic immune competent child. *J Child Neurol.* 2010;25(7):901-8.
4. Walwyn WM, et al. HSV-1-mediated NGF delivery delays nociceptive deficits in a genetic model of diabetic neuropathy. *ExpNeurol.* 2006;198(1):260-70.
5. Rainov NG and Heidecke V. Clinical development of experimental virus-mediated gene therapy for malignant glioma. *Anticancer Agents Med Chem.* 2011;11(8):739-47.
6. Yu DS, et al. Synthetic radiation-inducible promoters mediated HSV-TK/GCV gene therapy in the treatment of oral squamous cell carcinoma. *Oral Dis.* 2010;16(5):445-52.
7. Yao F, et al. Development of a regulatable oncolytic herpes simplex virus type 1 recombinant virus for tumor therapy *J Virol.* 2010;84(16):8163-71.
8. Kolodkin-Gal D, et al. Herpes simplex virus delivery to orthotopic rectal carcinoma results in an efficient and selective antitumor effect. *Gene Ther.* 2009;16(7):905-15.
9. Lin SF, et al. Synergy of a herpes oncolytic virus and paclitaxel for anaplastic thyroid cancer. *Clin Cancer Res.* 2008;14(5):1519-28.
10. Sung MW, et al. Intratumoral adenovirus-mediated suicide gene transfer for hepatic metastases from colorectal adenocarcinoma: results of a phase I clinical trial. *MolTher.* 2001;4(3):182-91.
11. Martins I, et al. Reversal of neuropathic pain by HSV-1-mediated decrease of noradrenaline in a pain facilitatory area of the brain *Pain.* 2010;151(1):137-45.
12. Wolfe D, et al. A clinical trial of gene therapy for chronic pain. *Pain Med.* 2009;10(7):1325-30.
13. Wolfe D, Mata M and Fink DJ. A human trial of HSV-mediated gene transfer for the treatment of chronic pain. *Gene Ther.* 2009;16(4):455-60.
14. Muhammad AK, et al. Study of the efficacy, bio distribution, and safety profile of therapeutic gutless adenovirus vectors as a prelude to a phase I clinical trial for glioblastoma. *Clin Pharmacol Ther.* 2010;88(2):204-13.
15. Rachakatla RS, et al. Combination treatment of human umbilical cord matrix stem cell-based interferon-beta gene therapy and 5-fluorouracil significantly reduces growth of metastatic human breast cancer in SCID mouse lungs *Cancer Invest.* 2008;26(7):662-70.
16. Kim KH, et al. A Phase I Clinical Trial of Ad5.SSTR/TK.RGD, a Novel Infectivity-Enhanced Bicistronic Adenovirus, in Patients with Recurrent Gynecologic Cancer. *Clin Cancer Res;* 2012.

17. Habib N, et al. Clinical trial of E1B-deleted adenovirus (dl1520) gene therapy for hepatocellular carcinoma. *Cancer Gene Ther.* 2002;9(3):254-9.
18. Rosengart TK, et al. Angiogenesis gene therapy: phase I assessment of direct intramyocardial administration of an adenovirus vector expressing VEGF121 cDNA to individuals with clinically significant severe coronary artery disease. *Circulation.* 1999;100(5):468-74.
19. Nayak S and Herzog RW. Progress and prospects: immune responses to viral vectors. *Gene Ther.* 2010;17(3):295-304.
20. Wu TL and Ertl HC. Immune barriers to successful gene therapy. *Trends Mol Med.* 2009;15(1):32-9.
21. Li C, et al. Cellular immune response to cryptic epitopes during therapeutic gene transfer. *Proc Natl Acad Sci, U S A.* 2009;106(26):10770-4.
22. Roelvink PW, et al. The coxsackievirus-adenovirus receptor protein can function as a cellular attachment protein for adenovirus serotypes from subgroups A, C, D, E, and F. *J Virol.* 1998;72(10):7909-15.
23. Tomko RP, Xu R and Philipson L. HCAR and MCAR: the human and mouse cellular receptors for subgroup C adenoviruses and group B coxsackieviruses. *Proc Natl Acad Sci, U S A* 1997;94(7):3352-6.
24. Raschperger E, et al. The coxsackie and adenovirus receptor (CAR) is an in vivo marker for epithelial tight junctions, with a potential role in regulating permeability and tissue homeostasis. *Exp Cell Res.* 2006;312(9):1566-80.
25. Chen S, et al. Ischemia post conditioning and mesenchymal stem cells engraftment synergistically attenuate ischemia reperfusion-induced lung injury in rats. *J Surg Res;* 2012.
26. Lundberg J, et al. Targeted intra-arterial transplantation of stem cells to the injured CNS is more effective than intravenous administration: engraftment is dependent on cell type and adhesion molecule expression. *Cell Transplant.* 2012;21(1):333-43.
27. Carney BJ and Shah K. Migration and fate of therapeutic stem cells in different brain disease models. *Neuroscience.* 2011;197:37-47.
28. Zhu F and Guo G. Research progress of bone marrow mesenchymal stem cells in acute lung injury. *Zhongguo Xiu Fu Chong JianWaiKeZaZhi.* 2011;25(2):198-201.
29. Mohsin S, et al. Enhanced hepatic differentiation of mesenchymal stem cells after pretreatment with injured liver tissue. *Differentiation.* 2011;81(1):42-8.
30. Fong EL, Chan CK and Goodman SB. Stem cell homing in musculoskeletal injury. *Biomaterials.* 2011;32(2):395-409.
31. Borlongan CV, et al. The great migration of bone marrow-derived stem cells toward the ischemic brain: therapeutic implications for stroke and other neurological disorders. *ProgNeurobiol.* 2011;95(2):213-28.
32. Asanuma H, Meldrum DR and Meldrum KK. Therapeutic applications of mesenchymal stem cells to repair kidney injury *J Urol.* 2010;184(1):26-33.
33. Doucette T, et al. Mesenchymal stem cells display tumor-specific tropism in an RCAS/Ntv-a glioma model. *Neoplasia.* 2011;13(8):716-25.
34. Croitoru-Lamoury J, et al. Human mesenchymal stem cells constitutively express chemokines and chemokine receptors that can be upregulated by cytokines, IFN-beta, and Copaxone. *J Interferon Cytokine Res.* 2007;27(1):53-64.
35. Ji JF, et al. Interactions of chemokines and chemokine receptors mediate the migration of mesenchymal stem cells to the impaired site in the brain after hypoglossal nerve injury. *Stem Cells.* 2004;22(3):415-27.
36. Carrero R, et al. IL1 $\beta$  induces mesenchymal stem cells migration and leucocyte chemotaxis through NF- $\kappa$ B. *Stem cell reviews.* 2012;8:905-16.

37. Anton K, Banerjee D and Glod J. Macrophage-associated mesenchymal stem cells assume an activated, migratory, pro-inflammatory phenotype with increased IL-6 and CXCL10 secretion. *PloS One*. 2012;7:35036.
38. Veevers-Lowe J, et al. Mesenchymal stem cell migration is regulated by fibronectin through  $\alpha 5\beta 1$ -integrin-mediated activation of PDGFR- $\beta$  and potentiation of growth factor signals. *Journal of Cell Science*. 2011;124:1288-300.
39. De Miguel MP, et al. Immunosuppressive properties of mesenchymal stem cells: advances and applications. *CurrMol Med*; 2012.
40. Menard C and Tarte K. Immunosuppression and mesenchymal stem cells: back to the future. *Med Sci (Paris)*. 2011;27(3):269-74.
41. Wang Q, et al. The allogeneic umbilical cord mesenchymal stem cells regulate the function of T helper 17 cells from patients with rheumatoid arthritis in an in vitro co-culture system. *BMC Musculoskeletal Disorders*. 2012;13:249.
42. Duffy MM, et al. Mesenchymal stem cell inhibition of T-helper 17 cell- differentiation is triggered by cell-cell contact and mediated by prostaglandin E2 via the EP4 receptor. *Eur J Immunol*. 2011;41(10):2840-51.
43. Ren G, et al. Inflammatory cytokine-induced intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 in mesenchymal stem cells are critical for immunosuppression. *J Immunol*. 2010;184(5):2321-8.
44. Najar M, et al. Modulated expression of adhesion molecules and galectin-1: role during mesenchymal stromal cell immunoregulatory functions. *ExpHematol*. 2010;38(10):922-32.
45. Du J, et al. IFN- $\gamma$ -primed human bone marrow mesenchymal stem cells induce tumor cell apoptosis in vitro via tumor necrosis factor-related apoptosis-inducing ligand. *The International Journal of Biochemistry and Cell Biology*. 2012;44:1305-14.
46. Zou W, et al. LIGHT delivery to tumors by mesenchymal stem cells mobilizes an effective antitumor immune response. *Cancer research*. 2012;72:2980-9.
47. Seo SH, et al. The effects of mesenchymal stem cells injected via different routes on modified IL-12-mediated antitumor activity. *Gene Ther*. 2011;18(5):488-95.
48. Zhao WH, et al. Human umbilical cord mesenchymal stem cells with adenovirus-mediated interleukin 12 gene transduction inhibits the growth of ovarian carcinoma cells both in vitro and in vivo]. *Nan Fang Yi Ke Da XueXueBao*. 2011;31(5):903-7.
49. Gao P, et al. Therapeutic potential of human mesenchymal stem cells producing IL-12 in a mouse xenograft model of renal cell carcinoma. *Cancer Lett*. 2010;290(2):157-66.
50. Hu W, et al. Human umbilical blood mononuclear cell-derived mesenchymal stem cells serve as interleukin-21 gene delivery vehicles for epithelial ovarian cancer therapy in nude mice. *Biotechnol Appl Biochem*. 2011;58(6):397-404.
51. Hu W, et al. Human umbilical blood mononuclear cell-derived mesenchymal stem cells serve as interleukin-21 gene delivery vehicles for epithelial ovarian cancer therapy in nude mice. *Biotechnology and Applied Biochemistry*. 58;397-404.
52. Wang Q, et al. Mesenchymal stem cells over-expressing PEDF decreased the angiogenesis of gliomas. *Bioscience reports*; 2012.
53. Chen Q, et al. Therapeutic potential of bone marrow-derived mesenchymal stem cells producing pigment epithelium-derived factor in lung carcinoma. *International journal of molecular medicine*. 2012;30:527-34.
54. Ryu CH, et al. Valproic acid enhances anti-tumor effect of mesenchymal stem cell mediated HSV-TK gene therapy in intracranial glioma. *BiochemBiophys Res Commun*; 2012.
55. Matuskova M, et al. HSV-tk expressing mesenchymal stem cells exert bystander effect on human glioblastoma cells. *Cancer Lett*. 2010;290(1):58-67.



56. Song C, et al. Thymidine kinase gene modified bone marrow mesenchymal stem cells as vehicles for antitumor therapy. *Hum Gene Ther.* 2011;22(4):439-49.
57. Fei S, et al. The antitumor effect of mesenchymal stem cells transduced with a lentiviral vector expressing cytosine deaminase in a rat glioma model. *J Cancer Res ClinOncol.* 2012;138(2):347-57.
58. Altanerova V, et al. Human adipose tissue-derived mesenchymal stem cells expressing yeast cytosinedeaminase: uracil phosphoribosyltransferase inhibit intracerebral rat glioblastoma. *Int J Cancer.* 2012;130(10):2455-63.
59. Chang DY, et al. The growth of brain tumors can be suppressed by multiple transplantation of mesenchymal stem cells expressing cytosine deaminase. *Int J Cancer.* 2010;127(8):1975-83.
60. Cavarretta IT, et al. Adipose tissue-derived mesenchymal stem cells expressing prodrug-converting enzyme inhibit human prostate tumor growth. *MolTher.* 2010;18(1):223-31.
61. You MH, et al. Cytosine deaminase-producing human mesenchymal stem cells mediate an antitumor effect in a mouse xenograft model. *J GastroenterolHepatol.* 2009;24(8):1393-400.
62. Kang NH, et al. Human amniotic fluid-derived stem cells expressing cytosine deaminase and thymidine kinase inhibits the growth of breast cancer cells in cellular and xenograft mouse models. *Cancer Gene Ther;* 2012.
63. Ryu CH, et al. Valproic acid enhances anti-tumor effect of mesenchymal stem cell mediated HSV-TK gene therapy in intracranial glioma. *Biochemical and Biophysical Research Communications.* 2012;421:585-90.
64. Cho YH, et al. Enhancement of MSC adhesion and therapeutic efficiency in ischemic heart using lentivirus delivery with periostin. *Biomaterials.* 2012;33(5):1376-85.
65. Zhang X, et al. Mesenchymal stem cells modified to express lentivirus TNF-alpha Tumstatin(45-132) inhibit the growth of prostate cancer. *J Cell Mol Med.* 2011;15(2):433-44.
66. Ghaedi M, et al. Mesenchymal stem cells as vehicles for targeted delivery of anti-angiogenic protein to solid tumors. *J Gene Med.* 2011;13(3):171-80.
67. Zou D, et al. Repair of critical-sized rat calvarial defects using genetically engineered bone marrow-derived mesenchymal stem cells overexpressing hypoxia-inducible factor-1alpha. *Stem Cells.* 2011;29(9):1380-90.
68. Ren G and Boison D. Engineering human mesenchymal stem cells to release adenosine using miRNA technology. *Methods Mol Biol.* 2010;650:225-40.
69. Gao Y, et al. Human mesenchymal stem cells overexpressing pigment epithelium-derived factor inhibit hepatocellular carcinoma in nude mice. *Oncogene.* 2010;29(19):2784-94.
70. Chong MS and Chan J. Lentiviral vector transduction of fetal mesenchymal stem cells. *Methods Mol Biol.* 2010;614:135-47.
71. Feng SW, et al. Restoration of muscle fibers and satellite cells after isogenic MSC transplantation with microdystrophin gene delivery. *BiochemBiophys Res Commun.* 2012;419(1):1-6.
72. Manning E, et al. Interleukin-10 delivery via mesenchymal stem cells: a novel gene therapy approach to prevent lung ischemia-reperfusion injury. *Hum Gene Ther.* 2010;21(6):713-27.
73. Okada T. Gene therapy with vector-producing multipotentmesenchymal stromal cells. *YakugakuZasshi.* 2010;130(11):1513-8.
74. Bobis-Wozowicz S, et al. Genetically modified adipose tissue-derived mesenchymal stem cells overexpressing CXCR4 display increased motility, invasiveness and homing to bone marrow of NOD/SCID mice. *ExpHematol.* 2011;39(6):686-6964.

75. Liu WW, et al. Hepatocyte growth factor combined with autologous bone marrow mesenchymal stem cell transplantation for treatment of silicosis. *Zhonghua Lao Dong Wei Sheng Zhi Ye Bing ZaZhi*. 2011;29(1):39-43.
76. Jiang YB, et al. Effects of heme oxygenase-1 gene modulated mesenchymal stem cells on vasculogenesis in ischemic swine hearts. *Chin Med J (Engl)*. 2011;124(3):401-7.
77. Ahmed AU, et al. Bone marrow mesenchymal stem cells loaded with an oncolytic adenovirus suppress the anti-adenoviral immune response in the cotton rat model. *Mol Ther*. 2010;18(10):1846-56.
78. Hamedi-Asl P, et al. Adenovirus-mediated expression of the HO-1 protein within MSCs decreased cytotoxicity and inhibited apoptosis induced by oxidative stresses. *Cell Stress Chaperones*. 2012;17(2):181-90.
79. Rasanen K and Herlyn M, Paracrine signaling between carcinoma cells and mesenchymal stem cells generates cancer stem cell niche via epithelial-mesenchymal transition. *Cancer Discov*. 2012;2(9):775-7.
80. Li HJ, et al. Cancer-stimulated mesenchymal stem cells create a carcinoma stem cell niche via prostaglandin E2 signaling. *Cancer Discov*. 2012;2(9):840-55.
81. Tsukamoto S, et al. Mesenchymal stem cells promote tumor engraftment and metastatic colonization in rat osteosarcoma model. *Int J Oncol*. 2012;40(1):163-9.
82. Szebeni GJ, et al. Identification of galectin-1 as a critical factor in function of mouse mesenchymal stromal cell-mediated tumor promotion. *PLoS One*. 2012;7:41372.
83. Ma M, et al. Mesenchymal stromal cells may enhance metastasis of neuroblastoma via SDF-1/CXCR4 and SDF-1/CXCR7 signaling. *Cancer Lett*. 2011;312(1):1-10.
84. Hsu HS, et al. Mesenchymal stem cells enhance lung cancer initiation through activation of IL-6/JAK2/STAT3 pathway. *Lung Cancer*. 2012;75(2):167-77.
85. Nakashima H, et al. Involvement of the transcription factor twist in phenotype alteration through epithelial-mesenchymal transition in lung cancer cells. *Mol Carcinog*. 2012;51(5):400-10.
86. Yan XL, et al. Mesenchymal stem cells from primary breast cancer tissue promotes cancer proliferation and enhances mammosphere formation partially via EGF/EGFR/Akt pathway. *Breast Cancer Res Treat*. 2012;132(1):153-64.
87. Djouad F, et al. Immunosuppressive effect of mesenchymal stem cells favors tumor growth in allogeneic animals *Blood*. 2003;102(10):3837-44.

© 2014 Nguyen et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Peer-review history:*

*The peer review history for this paper can be accessed here:*  
<http://www.sciencedomain.org/review-history.php?iid=397&id=32&aid=3301>