European Journal of Medicinal Plants



32(7): 19-29, 2021; Article no.EJMP.70422 ISSN: 2231-0894, NLM ID: 101583475

Hydro-Alcoholic Root Extracts of *Ziziphus abyssinica* is Effective in Diabetes Nephropathy and Diabetic Wound Healing

Samuel A. Akwetey¹, Douglas B. Aidoo^{2*}, Wisdom Ahlidja¹, Bright B. Boafo³, Joseph M. Acquah¹, Joshua F. Agyei¹, Desmond Bakoji¹ and Portia A. Okyir¹

¹Department of Biomedical Science, School of Allied Health Sciences, University of Cape Coast, Cape Coast, Ghana. ²Department of Pharmacology, Faculty of Pharmacy and Pharmaceutical Sciences, College of Health

²Department of Pharmacology, Faculty of Pharmacy and Pharmaceutical Sciences, College of Health Sciences, Kwame Nkrumah University of Science and Technology, KNUST, Kumasi, Ghana. ³Department of Toxicology, University of Saskatchewan, Saskatoon, SK, Canada.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/EJMP/2021/v32i730402 <u>Editor(s):</u> (1) Dr. Patrizia Diana, University of Palermo, Italy. (2) Dr. Daniela Rigano, University Federico II of Naples, Italy. (3) Prof. Marcello Iriti , Milan State University, Italy. <u>Reviewers:</u> (1) David Yakubu Bot, University of Jos, Nigeria. (2) Banu Buyukaydin, Bezmialem Vakif University, Turkey. Complete Peer review History: <u>https://www.sdiarticle4.com/review-history/70422</u>

Original Research Article

Received 23 May 2021 Accepted 29 July 2021 Published 17 August 2021

ABSTRACT

Background: This study evaluated the potential of *Ziziphus abysinnica* root extract in managing hyperglycaemia in type 2 diabetes mellitus (T2DM), diabetic wound healing and diabetic nephropathy.

Methodology: Blood glucose concentrations were measured daily for 14 days after daily administrations of either *Ziziphus abysinnica* (30, 100, and 300 mg/kg, *p.o*), metformin (300 mg/kg, *p.o*) or normal saline as negative control before diabetes induction using a single dose of Streptozotocin (60 mg/kg, i.p) and nicotinamide (120 mg/kg, i.p). Histopathological analysis was performed on the harvested kidneys following administration with *Ziziphus abysinnica* in diabetic rats. The diabetic wound healing potentials of the plant was also evaluated in streptozotocin-induced diabetic rats by treating them with 15%w/w ZAE ointment.

*Corresponding author: E-mail: mraidooucc@gmail.com;

Results: Generally, the percentage of blood glucose levels analysed following administration of drugs were found to be dose-dependent. The highest dose of *ZAE* (300 mg/kg) had a higher percentage reduction in blood glucose concentration when compared to metformin (300 mg/kg). The lowest dose (30 mg/kg) of ZAE administered attenuated STZ induced pathological damage and showed moderate to maximal improvement to the kidney nephrons. In contrast, the 100 mg/kg and 300 mg/kg dose ZAE demonstrated minimal pathological changes to the kidney architecture. **Conclusion:** Overall, our study demonstrated the antidiabetic potential of *Ziziphus abysinnica*, suggesting its possible therapeutic benefit in diabetic wound healing and diabetic nephropathy.

Keywords: Diabetic nephropathy; diabetic wound; type 2 Diabetes mellitus; Ziziphus abysinnica.

1. BACKGROUND

Diabetes mellitus (DM) is a metabolic disease characterised by a relative or absolute lack of insulin. resultina in hyperglycaemia [1]. Persistent DM can lead to chronic hyperglycaemia, which leads to a variety of multiorgan complications such as neuropathy, nephropathy, retinopathy, increased risk of cardiovascular diseases, and impaired wound healing. The wound healing impairment observed in diabetes could be caused by numerous factors such as inadequate blood supply, reduced proliferation of fibroblast, and decreased inflammatory changes [2]. The International Diabetes Federation estimates that currently, 463 million adults worldwide are affected by DM, and this figure is expected to reach a staggering 578 million by 2030 [3], of which type 2 diabetes (T2D) accounts for more than 90 % of the cases [4]. T2D is a complex, heterogeneous and polygenic disease characterised mainly by insulin resistance and pancreatic β -cell dysfunction [5], which leads to chronic hyperglycemia. Diabetes is though considered one of the five leading causes of mortality worldwide, however, no satisfactory effective therapy is available to cure diabetes in modern medicine. Currently, T2D is predominantly managed with oral antidiabetic drugs such as sulfonylureas, biguanides, and α glucosidase inhibitors [6]. Unfortunately, these synthetic antidiabetic agents are beset with some side effects such as diarrhoea, nausea, and liver failure [7].

The kidneys are one of the primary targets, which are affected by diabetes mellitus (DM). Diabetic nephropathy (DN) is the leading cause of chronic kidney failure, and consequently, a significant cause of renal morbidity and mortality [8]. It is estimated that about 40% of all people with DM develop clinical evidence of nephropathy, but a considerably smaller fraction of type II diabetic patients' progress to end-stage renal disease (ESRD). Advanced or end-stage kidney disease occurs in as many as 40 % of both type I and type II diabetics [9,10].

Ziziphus abyssinica is a semi-deciduous flowering shrub or tree which belongs to the family Rhamnaceae. The plant has a widespread distribution almost throughout Africa [11] and is commonly known as catch thorn. A decoction of the plant is traditionally used to treat diabetes mellitus [12] among the rural inhabitants of northern Nigeria and some parts of Ghana. The roots are boiled, and the liquid is drunk as a treatment for early postpartum pains, stomach aches, snake bites, and an abortifacient. The leaves are also boiled and used as a steam bath to treat pneumonia [13]. The extract from the plant's edible fruit has been reported to possess antioxidant properties and antibacterial activity against Pseudomonas aeruginosa, Escherichia coli. Staphylococcus aureus, and Candida albicans [14]. The root extract of the plant is known to possess anti-plasmodial [15], antiulcerative [16], analgesic [17], and anti-diarrhoeal properties [18]. Additionally, the plant's root back extract has been reported to possess an inhibitory effect against acute inflammation [19].

Despite the widespread use of the plant in folklore medicine across many African countries, no scientific data is backing this plant's roots in managing diabetes and its associated complications such as diabetic nephropathy. This study hypothesized that Ziziphus abyssinica has a blood glucose-reducing effect and heals wounds in diabetic rats.

2. METHODS

2.1 Plant Material

The roots of *Ziziphus abyssinica* were collected from Ejura, in the Ashanti region of Ghana, in June 2017 and authenticated by a botanist (Mr Clifford Asare) at the Herbarium Unit of Faculty of Pharmacy and Pharmaceutical Sciences, Kwame Nkrumah University of Science and Technology (KNUST). The roots were shadedried for one month and were compared with a voucher specimen KNUST/HM/2016/L003 kept in the herbarium.

2.2 Plant Extraction

The dried roots were pulverized into powder with an electric mill. One (1) kg of the powdered material was soaked with 3 L of 70 % v/v ethanol and maintained on a mechanical shaker for 72 h. The extract obtained was filtered and labelled ZAE and the filtrate concentrated using a rotary evaporator (Rotavapor R-215 model, BÜCHI Labortechnik AG, Flawil, Switzerland) under reduced pressure (50 °C) and temperature (40 °C). The filtrate was then placed in a desiccator containing activated silica gel to dry, and the dried mass was refrigerated at 4 °C until ready to be used.

2.3 Preparation of Ointment

Formulation of ointment was done using a method as described [20] with modification. The ointment was prepared by taking 0.3 % preservatives (methylparaben, propylparaben), 30 % humectants (petroleum jelly), and 19.5 % emulsifying wax in 500 ml of distilled water and 1.5 % emulsifying agent (acetyl alcohol), and 15 % glycerin in 45 % liquid paraffin. Both oily and aqueous phases were mixed at 700 °C, and three graded fractions of the cooled extract were added separately. The resultant ointment was homogenized by Soxhlet, a homogenizer at 3000 rpm, into a creamy form and stored in tight plastic containers.

2.4 Animals

Male Sprague-Dawley (SD) rats (250 - 300 g) were purchased from the Noguchi Memorial Institute for Medical Research, University of Ghana, Ghana. The animals were housed in stainless steel cages (34 x 47 x 18) cm³ in groups of six rats at the animal house facility of the Department of Biomedical Sciences, University of Cape Coast, Ghana. The male SD rats were maintained under standard laboratory conditions, 12-hour light/dark cycle, standard food and water ad libitum, 25 °C, and free movement in their cages. Ethical approval for the use of experimental animals was approved by the Ethics Review Committee of the University of Board Coast Institutional Review Cape (UCCIRB). UCCIRB/CHAS/2016/13. ID:

Experimental animals were handled per the Animal Welfare Regulation and the Public Health Service Policy on Humane Care and Use of Laboratory animals [21].

2.5 Drugs and Chemicals

Streptozotocin (STZ) and nicotinamide were procured from (Sigma-Aldrich Inc, St. Louis, MO, USA). Metformin hydrochloride and becaplermin ointment was acquired from (Ernest Chemist, Accra, Ghana).

2.6 Experimental Design

2.6.1 Induction of diabetes

Diabetes was induced in overnight fasted rats following a described method of [22] with slight modification. Sprague-Dawley rats (250 - 300 g) were randomized into six groups (n=5) and given one of the following treatments for 14 days post-induction.

Group I (naïve control): normal saline 10 ml/kg, p.o.

Group II (positive control): metformin 300 mg/kg, p.o + becaplemin ointment

Group III: (disease control): normal saline 10 ml/kg, *i.p.*

Groups IV, V, and VI (test groups): *Ziziphus abysinnica* (ZAE) 30, 100, and 300 mg/kg p.o. respectively.

Test rats (except group I) received a single intraperitoneal injection (*i.p.*) of nicotinamide (120 mg/kg dissolved in normal saline). After 20 min post nicotinamide administration, test animals were given (60 mg/kg, i.p.) Streptozotocin (STZ) dissolved in 2 ml/kg citrate buffer (pH= 4.5).

2.6.2Confirmation of type 2 diabetes mellitus (T2DM)

Fasting blood glucose (FBG) concentration was measured using samples from the lateral tail vein with the help of a glucometer (OneTouch® Ultra, USA). Experimental animals with fasting blood glucose levels of 10 mmol/L or greater were considered diabetic and were used for the study.

2.6.3 Blood glucose monitoring

Blood glucose concentrations were measured after an overnight fast using a glucometer. Small venipuncture was made on the lateral part of the tail of each animal, and blood was obtained for measuring blood glucose levels. Twelve (12) hours after the last treatment, the fasting blood sugar (FBS) levels were measured again.

2.6.4 Excision wound model

The rats were anaesthetized with slight vapour inhalation of diethyl ether. The hair at the vertebral area of the rats was shaved before wound creation, and the application field for the ointment was outlined with a marking pen. An excision wound of 1.5cm in diameter and 2mm depth was created on the dorsum of the rats using sterile toothed forceps and surgical blades.

2.6.5 The wound healing evaluation

Screening for the wound healing activity of the ointment was performed by the excision wound model as described [23]. The diabetic animals were divided into 5 sub-groups, each containing 5 animals. The control group (group I) was treated topically with sterile normal saline. Groups IV, V, and VI were treated topically with ZAE ointment (15 % w/w), whereas group II received topical application of Becaplemin gel 0.01 % (Regranex) for 14 days. The progressive changes in wound area were measured in mm² by tracing the wound boundaries around it on transparent polythene paper daily for 14 days of treatment. The polythene paper was then placed on the graph paper and traced out. The healed area was calculated by subtracting from the total wound area as follows:

```
% wound closure = \frac{(initial area of wound-nth day area of wound)}{(initial area of wound)} x 100
```

2.6.6. Histology of kidney tissue

Experimental animals were euthanised through anaesthesia, and their kidneys were carefully harvested and preserved in 10 % neutered buffered formalin at room temperature for microscopic evaluations. The harvested kidney tissues were fixed in 10 % formalin, dehydrated in 70 % ethanol solutions of increasing concentrations (50-100 %), cleared in xylene, and embedded in paraffin wax. Transverse sections of 5 µm were obtained using a microtome (Bright 5040, Bright Instrument Company Ltd., England) and stained with hematoxylin and eosin (H&E) for microscopic evaluation of tissue integrity. Microscopic examinations were done using a binocular clinical light microscope with a digital camera (Olympus CX1, Japan) connected to a computer. Photomicrographs of the tissues were generated

using the ×100 objective lens for further analysis of pathological abnormalities such as leukocyte infiltration, haemorrhage, fatty changes, necrosis, and congestion.

2.7 Statistical Analysis

GraphPad Prism for Windows Version 7.0 (GraphPad Software, San Diego, CA, USA, 2016) was used for all statistical analyses. Data were expressed as a mean \pm standard error of the mean (SEM). Differences among treatment group means were assessed using a one-way analysis of variance (ANOVA) followed with the Dunnett post hoc test. P < 0.05 was considered statistically significant for all tests.

3. RESULTS

3.1 Effect of *Ziziphus abysinnica* on the blood glucose levels

The blood glucose concentrations were measured in normal and experimental rats on days 2, 4, 6, 8, 10, 12, and 14 of treatment (Fig. 1). The percentage reduction of blood glucose in the treatment groups increased steadily from day 2, with the 300 mg/kg ZAE treatment group having the highest value (% reduction) by day 14. ZAE treatment groups (groups V - VI) showed a dose-dependent hypoglycaemic effect compared to the naïve control (group I). Metformin (300 mg/kg) showed a higher percentage reduction when compared to ZAE (30 mg/kg, 100mg/kg) respectively (Fig. 2).

3.2 Wound healing effect of *Ziziphus abyssinica* extract ointment in streptozotocin-induced diabetes

ZAE ointment demonstrated a significant total wound healing percentage reduction of 2.5 %, 5 %, and 10 % respectively compared to the negative control (p<0.005) beginning from the 8th day through to the 14th day (Fig. 3). ZAE offered a mean coverage of 705.0±6.163, 679.7±37.62, and 690±32.69 rate of wound contraction compared to the negative control (515.9±52.63). The becaplermin ointment had total mean coverage of 684.2±26.62 at a significance of p<0.05 against the negative control (Fig. 4)

3.3 Effect Ziziphus abysinnica on kidney histology

Plate 1 presents kidney micrographs of naïve control rats, diabetic rats with no treatments, and rats that received metformin, the various doses

of the ZAE plant extracts. No morphological changes were observed in saline control rats (Plate 1 GI). Diabetic rats without treatment show severe kidney damage in the form of increased capsular space, vasodilatations, and tubular hypertrophy (Plate 1 GIII). Treated SD rats with metformin show severe thickening of the basement membrane of glomeruli alongside moderate changes, including congestion and shrinkage of glomerular (Plate 1 GII). Diabetic rats treated with the least dose (30 mg/kg) of extract showed mild pathological changes except for shrinkage of glomerular, which was low (Plate 1 GV). Meanwhile, the 100 mg/kg and 300mg/kg ZAE treatment groups ameliorated the severe pathological changes and improved the thickening of the basement membrane of glomeruli (Plate 1 GIV and GVI).

4. DISCUSSION

Results obtained after the 14-day intervention showed that the Ziziphus abvssinica extract effectively lowered blood alucose concentration at all doses in a dose-dependent manner. The highest dose of the extract (300 mg/kg) had the most profound antidiabetic effect, reducing blood glucose concentration by nearly 60 % followed by 100 mg/kg (30 %) and 30 mg/kg (28 %) of the plant extract respectively. From the results, it can be seen that the higher the dose(s) of the plant extract, the higher the percentage reduction and vice versa. Thus, the anti-hyperglycaemic activity of the plant extract exhibits a dose-dependent effect. Again, from the results, the comparison of the mean fasting blood glucose concentration values of graded doses of the plant extract with

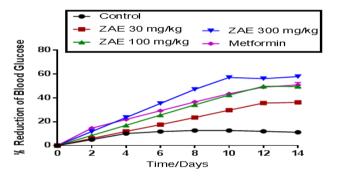


Fig. 1. Effect of *Ziziphus abyssinica* on percentage reduction of blood glucose level in STZinduced diabetic rats; Sprague-Dawley rats received either saline 10 ml/kg, metformin 300 mg/kg, or *Ziziphus abysinnica* 30, 100, and 300 mg/kg p.o., and challenged with streptozotocin (60 mg/kg, i.p.) 20 min post-administration of nicotinamide (120 mg/kg i.p.). Fasting blood glucose concentration was measured before and 12 h after treatments

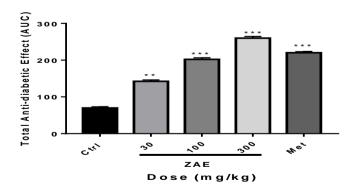


Fig. 2. Effect of *Ziziphus abyssinica* on the mean fasting blood glucose concentration in STZinduced diabetic rats; Sprague-Dawley rats received either saline 10 ml/kg, metformin 300 mg, or *Ziziphus abysinnica* 30, 100, and 300 mg/kg p.o. Test SD rats were challenged with streptozotocin (60 mg/kg, i.p.) 20 min post-administration of nicotinamide (120 mg/kg i.p.). Fasting blood glucose concentration was measured before and 12 h after treatments

Akwetey et al.; EJMP, 32(7): 19-29, 2021; Article no.EJMP.70422

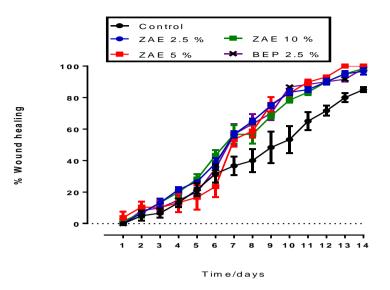


Fig. 3. Assessment of *Ziziphus abyssinica* ointment on wound healing (% reduction in wound diameter) in STZ-induced diabetic rats treated topically with 2.5 % becaplermin ointment or 2.5 %, 5.0 %, and 10 % *Ziziphus abyssinica*

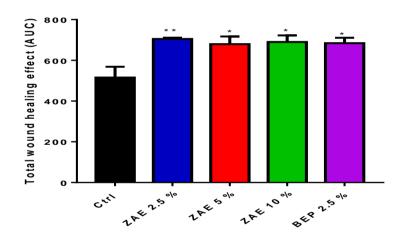


Fig. 4. Assessment of Ziziphus abyssinica ointment on total mean wound healing in STZinduced diabetic rats treated topically with 2.5% becaplermin ointment, or 2.5%, 5.0%, and 10% of Ziziphus abyssinica ointment for 14 days. ** p< 0.005 and *p<0.01

that of the diabetic control group all gave a pvalue which was less than 0.05. This means that the mean fasting blood glucose concentration was significantly less in treated SD rats when compared to the diabetic control group.

The anti-hyperglycaemic activity exhibited by the plant extract in this experiment can be attributed to the bioactive compounds present in the plant. Previous studies on the phytochemical analysis of *Ziziphus abyssinica* showed that the plant contains alkaloids, tannins, flavonoids, saponins, and steroids [14]. These active constituents

(alkaloids, tannins, flavonoids, and steroids) are all families of compounds that have been documented to possess an anti-hyperglycaemic effect [24]. Thus, one or a combination of some or all of the compounds mentioned above could responsible have been for the antihyperglycaemic effect of the extract. Highlighting the exact mechanism of action of some of these compounds, [25] reported that saponins act as the anti-hyperglycemic agent by lowering glucagon levels which were evidenced by a decrease in glucose-6-phosphatase and fructose 1, 6-diphosphatase activities. Both glucose-6-

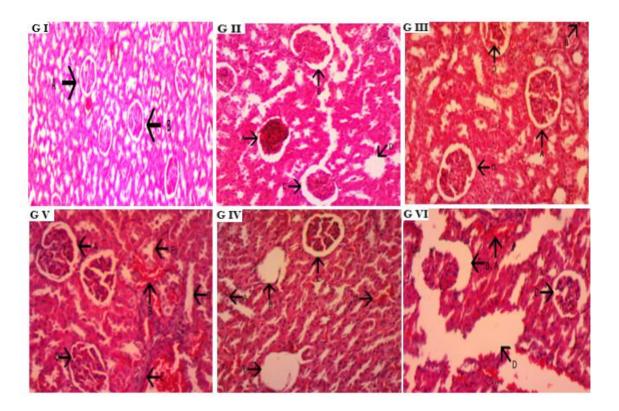


Plate 1. Photomicrograph of Ziziphus abysinnica on kidney damage in STZ-induced diabetes Sprague-Dawley rats received either saline 10 ml/kg, metformin 300 mg/kg, or Ziziphus abysinnica 30, 100, and 300 mg/kg p.o. Test SD rats were challenged with streptozotocin (60 mg/kg, i.p.) 20 min post-administration of nicotinamide (120 mg/kg i.p.). Rats were sacrificed, kidneys removed and fixed on 10% formalin, and embedded in paraffin. GI=normal control group. GII=positive control group (metformin 300 mg/kg (p.o). GIII= negative control group. Kidney micrographs for GV, GIV, and GVI= diabetes treated with 30, 100, and 300 mg/kg (p.o) ZAE extracts respectively. A, B, and C are sections of the kidney showing the renal corpuscles and renal tubules. A shows a normal architecture of the tubules and renal corpuscles whiles B and C show pathologic changes such as an increase in capsular space, tubular hypertrophy, thickening of the basement membrane of the glomeruli, vasodilatations, congestion, and shrinkage of the glomeruli in varying degrees

phosphatase and fructose 1, 6-diphosphatase essential enzvmes that are catalvse gluconeogenic reactions in the liver. The phosphorylation and activation of these enzymes are dependent on circulating levels of glucagon. In diabetes mellitus, there is uncontrolled hepatic gluconeogenesis, which contributes to hyperglycemia [26]. Thus, it could be possible that in diabetic animals, the lowering of glucagon levels by saponin inhibited gluconeogenesis in the liver, which consequently resulted in a decrease in the uncontrolled hepatic output of glucose into the plasma. Furthermore, tannins have also been reported to inhibit insulin degradation and improve glucose utilisation [27, 28].

Another mechanism by which the plant extract can cause its anti-hyperglycemic action may be through the stimulation of insulin secretion from the remaining intact pancreatic beta cells. This mechanism is postulated on the basis that the activity of the streptozotocin may have destroyed many pancreatic beta cells. Consequently, it is expected that the remaining pancreatic beta cells have to be stimulated to secrete a sufficient amount of insulin capable of causing a significant fall in the blood glucose level. As stated previously, the phytochemical screening of *Ziziphus abyssinica* revealed the presence of saponins which have been reported to stimulate insulin release from the pancreas [29]. Thus, saponins in the plant extract make the above-postulated mechanism more likely to be true.

In previous studies, their results showed a significant increase in serum levels of insulin in diabetic animals that were treated with *Ziziphus* extract [30, 31]. This observation indicates that Ziziphus extracts may enhance insulin release from pancreatic beta cells, either by regenerating the partially destroyed pancreatic beta cells or

releasing insulin stored in the granules. Furthermore, saponins have also been reported to inhibit the absorption of glucose from the intestine [32]. This saponin ability could be one of the alternative mechanisms through which the plant extract reduced the serum glucose concentrations in diabetic animals.

The normal control group (Plate 1 GI) showed regular cellular glomerular tufts in a tubules background with cuboidal cell epithelial lining with no pathological changes. In diabetes without treatment (Plate 1 GIII), the polyuria in the diabetic rats induced stress on the kidney cells, which caused congestion as a result of rupture of renal vessels and other pathological problems `such as tubular hypertrophy as a result of engorgement of tubules with fluids due to imbalance in osmolality caused by the damage to the nephrons [33].

Also, hyperglycaemia by itself is an independent risk factor for acute tubular injury due to the activation of free radicals and oxidative stress in tubular cells. In plate 1 GII, where the condition was treated with a standard drug (300 mg/kg of metformin), the kidney exhibited some moderate pathologic recovery such as thickening of the basement membrane of glomeruli which resulted from a loss of functionality of the epithelial cells and connective tissue cells to synthesise collagen fibres and reticular fibres. This consequently results in renal damage through renal hypoperfusion or endothelial injury through the release of various circulating substances. The lowest dose (30mg/kg) of the extract produced minimal or moderate pathological recovery to the kidney architecture, Treatment with ZAE 100mg/kg and 300mg/kg ameliorated the severe pathological damage and tubular hypertrophy which resulted from the loss of functionality of the epithelial cells and connective tissue cells to synthesise collagen fibres and reticular fibres and engorgement of tubules with fluids due to an imbalance in osmolality caused by the damage to the nephrons. Congestion of the tubules was because of damage to the renal veins caused by the toxic doses of the plant extract. This consequently led to the rapture of renal veins and engorgement of glomerular vessels with blood.

The study showed that topical application of ZAE ointment to diabetic wounds exhibited significant improvement in wound contraction and epithelialization on day eight and beyond compared to the control group. Wound

contraction helps the edges of the wound pull together, and epithelization causes new covering over the wound. ZAE ointment treated wounds were dry and uninflamed, indicating that ZAE roots contain abundant astringent and antiinflammatory properties. The effects observed could be attributed to the presence of tannins, flavonoids. and alkaloids [14]. Tannins possess astringent property. Alkaloids and flavonoids have also been shown to have anti-Flavonoids inflammatorv activity. also demonstrated high antioxidant and free radicalscavenging properties, enhancing the level of antioxidant enzymes in granuloma tissue [34].

Wound exudates usually provide a conducive environment for microbial growth. Bacteria such as P. aeruginosa and S. aureus isolated from wounds have been resistant to some antibacterial agents. The study observed that ZAE ointment could have promoted wound healing by inhibiting the growth of bacteria, resulting in possible wound healing. The antimicrobial activity may be due to the presence flavonoids [35] in ZAE. Sterols and of polyphenols have been reported to be responsible for wound healing due to their free radical-scavenging and antioxidant activity. These phytochemicals are also known to possess lipid peroxidation reduction abilities, which prevents cell necrosis and improves vascularitv (angiogenesis), thus increasing circulation (oxygen and essential nutrients) to the injured site [36]. The resultant effect is enhanced epithelial cell proliferation [37].

5. CONCLUSION

present Conclusively. the studv has demonstrated dose-dependent а antihyperglycemic and nephroprotective effect of the root extract of Ziziphus abyssinica (ZAE) in murine models. Even though the percentage glucose concentration reduction was much higher than metformin, there were moderate pathological changes to the kidneys. Topical application of ZAE ointment showed efficacy for treating and healing diabetic wounds.

ETHICAL APPROVAL

Ethical approval for the use of experimental animals was approved by the Ethics Review Committee of the University of Cape Coast Institutional Review Board (UCCIRB). ID: UCCIRB/CHAS/2016/13.

ACKNOWLEDGEMENT

We wish to express our gratitude to Mr Daniel Konja and Dr Isaac Tabiri Henneh for their immense contribution. The authors are grateful to the staff at the Department of Biomedical Sciences, University of Cape Coast

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Oršolić N, Bašić I. Honey bee products and their polyphenolic compounds in treatment of diabetes. Phytopharmacology and therapeutic values IV. 2008;443-540.
- Apikoglu- Rabus S, Izzettin FV, Turan P, Ercan FE. Effect of topical insulin on cutaneous wound healing in rats with or without acute diabetes. Clinical and Experimental Dermatology: Clinical dermatology. 2010;35(2):180-5.
- 3. Atlas D. International diabetes federation. IDF Diabetes Atlas, 7th edn. Brussels, Belgium: International Diabetes Federation;2015.
- Melmed S, Casanueva FF, Hoffman AR, Kleinberg DL, Montori VM, Schlechte JA, Wass JA. Diagnosis and treatment of hyperprolactinemia: an Endocrine Society clinical practice guideline. The Journal of Clinical Endocrinology & Metabolism. 2011;96(2):273-88.
- 5. Salas-Salvadó J, Bulló M, Babio N, Martínez-González MÁ, Ibarrola-Jurado N, Basora J, Estruch R, Covas MI, Corella D, Arós F. Ruiz-Gutiérrez V. Reduction in the incidence of type 2 diabetes with the diet: Mediterranean results of the PREDIMED-Reus nutrition intervention randomized trial. Diabetes care. 2011;34(1):14-9.
- Ghadyale V, Takalikar S, Haldavnekar V, Arvindekar A. Effective control of postprandial glucose level through inhibition of intestinal alpha glucosidase by Cymbopogon martinii (Roxb.). Evidence-Based Complementary and Alternative Medicine. 2012;2012.
- Fujisawa T, Ikegami H, Inoue K, Kawabata Y, Ogihara T. Effect of two α-glucosidase inhibitors, voglibose and acarbose, on

postprandial hyperglycemia correlates with subjective abdominal symptoms. Metabolism. 2005;54(3):387-90.

- Miranda-Díaz AG, Pazarín-Villaseñor L, Yanowsky-Escatell FG, Andrade-Sierra J. Oxidative stress in diabetic nephropathy with early chronic kidney disease. Journal of diabetes research. 2016;2016.
- Mora Fernández C, Domínguez Pimentel 9. V, de Fuentes MM, Górriz JL, Martínez Castelao A, Navarro González Diabetic kidney disease: from JF. physiology to therapeutics. The Journal of physiology. 2014;592(18):3997-4012.
- 10. Liu WJ, Huang WF, Ye L, Chen RH, Yang C, Wu HL, Pan QJ, Liu HF. The activity and role of autophagy in the pathogenesis of diabetic nephropathy. Eur Rev Med Pharmacol Sci. 2018;22(10):3182-89.
- Kaleem WA, Muhammad N, Khan H, Rauf A. Pharmacological and phytochemical studies of genus Zizyphus. Middle-East J Sci Res. 2014;21(8):1243-63.
- 12. Etuk EU, Bello SO, Isezuo SA, Mohammed BJ. Ethnobotanical survey of medicinal plants used for the treatment of Diabetes mellitus in the north western region of Nigeria. Asian Journal of Experimental Biological Sciences. 2010;1(1):55-9.
- Salih RR, Gibreel HH, Ahmed DM, Hammad ZM, Ahmed AI. Antimicrobial Effect of Aqueous and Ethanolic Leaves Extracts of Ziziphus Species against Animal Bacterial Pathogens. Curr Trends Forest Res. 2019;3(1036):2638-0013.
- 14. Nyaberi MO, Onyango CA, Mathooko FM, Maina JM, Makobe M, Mwaura F. Evaluation of phytochemical, antioxidant and antibacterial activity of edible fruit extracts of Ziziphus abyssinica A. Rich;2010.
- Muthaura CN, Keriko JM, Mutai C, Yenesew A, Gathirwa JW, Irungu BN, Nyangacha R, Mungai GM, Derese S. Antiplasmodial potential of traditional phytotherapy of some remedies used in treatment of malaria in Meru–Tharaka Nithi County of Kenya. Journal of Ethnopharmacology. 2015;175:315-23.
- Ugwah MO, Etuk EU, Bello SO, Aliero AA, Ugwah-Oguejiofor CJ. Comparative studies of anti-ulcerogenic activities of three Nigerian medicinal plants: a preliminary evaluation. Journal of Medicinal Plants Research. 2013;7(9):490-5.

- Boakye-Gyasi E, Henneh IT, Abotsi WK, Ameyaw EO, Woode E. Hydro-ethanolic leaf extract of Ziziphus abyssinica Hochst Ex A. Rich (Rhamnaceae) exhibits antinociceptive effects in murine models. BMC complementary and alternative medicine. 2017;17(1):1-2.
- Ugwah-Oguejiofor JC, Alkali IY, Ugwah MO, Abubakar K. Antidiarrhoeal potential of the aqueous root extract of Ziziphus abyssinica a. Rich, Sch Acad J Pharm. 2013;2:419-23.
- 19. Henneh IT, Ameyaw EO, Biney RP, Armah FA, Obese E, Konjah D, Teye E. Ziziphus abyssinica hydro-ethanolic root bark extract attenuates acute inflammation possibly through membrane stabilization and inhibition of protein denaturation and neutrophil degranulation. West African Journal of Pharmacy. 2018;29(2):81-94.
- 20. Chopda MZ, Nemade NV, Mahajan RT. Wound healing activity of root of Ziziphus jujuba mill in rat model. World Journal of Pharmacy and Pharmaceutical Sciences (WJPPS). 2014;3(9):830-6.
- Bailoo JD, Reichlin TS, Würbel H. Refinement of experimental design and conduct in laboratory animal research. ILAR journal. 2014 Dec 20;55(3):383-91.
- 22. Masiello P, Broca C, Gross R, Roye M, Manteghetti M, Hillaire-Buys D, Novelli M, Ribes G. Experimental NIDDM: development of a new model in adult rats administered streptozotocin and nicotinamide. Diabetes. 1998;47(2):224-9.
- 23. Ozay Y, Guzel-Ozay S, Cimbiz A, Gurlek-Olgun E, Ozyurt S. Effects of Equisetum arvense Ointment on Diabetic Wound Healing in Rats. Wounds: a compendium of clinical research and practice. 2013;25(9):234-41.
- 24. Fatima A, Singh PP, Irchhaiya R, Agarwal P. Effect of Leaves of Carissa spinarum Linn. on Blood Glucose and Lipid Profile in Alloxan Induced Diabetic Rats.24. Jamshidi HR, Mosaddegh MH.
- Vahidi AR, Ghasemian M, Haj Mohammadi N. The Effect of Ziziphus Jujuba Fruit Extract in Diabetic and Non-Diabetic Rat. Iran J diabetes Obes. 2014;6(1): 34–40.
- 26. Rang HP, Dale MM. Rang and Dale's pharmacology. Elsevier Brasil; 2007.

- 27. Marles RJ, Farnsworth NR. Antidiabetic plants and their active constituents. Phytomedicine. 1995;2(2):137-89.
- Peungvicha P, Thirawarapan SS, Temsiririrkkul R, Watanabe H, Prasain JK, Kadota S. Hypoglycemic effect of the water extract of Piper sarmentosum in rats. Journal of ethnopharmacology. 1998 ;60(1):27-32.
- Norberg A, Hoa NK, Liepinsh E, Van Phan D, Thuan ND, Jörnvall H, Sillard R, Östenson CG. A novel insulin-releasing substance, phanoside, from the plant Gynostemma pentaphyllum. Journal of Biological Chemistry. 2004; 279 (40):41361-7.
- 30. Glombitza KW, Mahran GH, Mirhom YW, Michel KG, Motawi TK. Hypoglycemic and antihyperglycemic effects of Zizyphus spina-christi in rats. Planta medica. 1994; 60 (03):244-7.
- Dash LA, Comstock GW, Flynn JP. Isoniazid preventive therapy: retrospect and prospect. American Review of Respiratory Disease. 1980; 121 (6):1039-44.
- Matsuda H, Li Y, Yamahara J, Yoshikawa M. Inhibition of gastric emptying by triterpene saponin, momordin Ic, in mice: roles of blood glucose, capsaicin-sensitive sensory nerves, and central nervous system. Journal of Pharmacology and Experimental Therapeutics. 1999; 289 (2):729-34.
- Tujios S, Fontana RJ. Mechanisms of drug-induced liver injury: from bedside to bench. Nature reviews Gastroenterology & hepatology. 2011;8 (4):202-11.
- 34. Shenoy C, Patil MB, Kumar R, Patil S. Preliminary phytochemical investigation and wound healing activity of Allium cepa Linn (Liliaceae). International Journal of Pharmacy and Pharmaceutical Sciences. 2009;2(2):167-75.
- Ayaz FA, Hayırlıoglu-Ayaz S, Alpay-Karaoglu S, Grúz J, Valentová K, Ulrichová J, Strnad M. Phenolic acid contents of kale (Brassica oleraceae L. var. acephala DC.) extracts and their antioxidant and antibacterial activities. Food Chemistry. 2008;107(1):19-25.
- 36. Baravkar AA, Kale RN, Patil RN, Sawant SD. Pharmaceutical and biological evaluation of formulated cream of

Akwetey et al.; EJMP, 32(7): 19-29, 2021; Article no.EJMP.70422

methanolic extract of Acacia nilotica leaves. Research Journal of Pharmacy and Technology. 2008;1(4):480-3. 37. Thakur R, Jain N, Pathak R, Sandhu SS. Practices in wound healing studies of plants. Evidence-based complementary and alternative medicine. 2011;2011.

© 2021 Akwetey et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle4.com/review-history/70422