



A Detailed Analysis of the Consequences of Various Nanoparticles on Growth, Development and Physiological Responses in Plants under Changing Environments

**Zabeehullah Burhan ^a, Sama Usman ^a, Hina Nazir ^a,
Narmeen Ayesha ^a, Areej Zubair ^b, Aliza Fermaish Ali ^{c*},
Saima Nadir Ali ^c and Kiran Fatima ^d**

^a Department of Botany, University of Agriculture Faisalabad, Pakistan.

^b Department Natural Sciences and Humanities Department, UET Lahore University, University of Engineering and Technology, New Campus, UET Lahore, Pakistan.

^c Department of Botany, University of Education Lahore, Pakistan.

^d Department of Chemistry, University of Agriculture Faisalabad, Pakistan.

Authors' contributions

This work was carried out in collaboration among all authors. All authors contributed to the study's conception and design. Author AFA was responsible for creating the study and writing the protocol. Authors AFA and ZB handled the preparation of the materials, data collection, and analysis. Author AFA wrote the first draft of the manuscript, and author SU provided feedback on earlier iterations. Authors HN, NA and AZ the literature searches and contributed a lot to Strategies Portion. The final part of the manuscript is Hinder Hunger written by authors SNA and HN. Author AFA was in charge of managing the references and citations. All authors read and approved the final manuscript.

Article Information

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/113997>

Review Article

Received: 04/01/2024

Accepted: 07/03/2024

Published: 18/03/2024

*Corresponding author: E-mail: authoralizainbotany93@gmail.com;

ABSTRACT

Enhancing plant nutrition without changing soil texture and protecting it from microbial diseases, nano-fertilizers, nano-pesticides, and nano-herbicides are some examples of how nanotechnology is being used in agriculture. So, nanotechnology keeps the soil healthy, which in turn keeps the plant healthy. Nanoparticles (NPs) increase agricultural productivity and production while decreasing chemical runoff and nutrient loss. Concentrations, physicochemical characteristics, and plant species all have a role in how NPs affect plants. There are a number of NPs that affect plant physiology, which in turn increases biomass production and germination rate. Meanwhile, the function of NPs in growth suppression, inhibition of chlorophyll, and photosynthetic efficiency has been extensively studied. To fill this review, we tried to compile studies that looked at NP effects, translocation, and interactions with plants. Also discussed are methods for phytoremediation of polluted soil that make use of NPs in conjunction with one another to promote environmentally responsible farming.

Keywords: Gene expression; nanotechnology; photosynthetic efficiency; phytoremediation; quantum dots.

1. INTRODUCTION

During the techno-science period, nanotechnology was at the forefront of innovation, drawing interest from many fields and industries that are directly related to human well-being, such as plant and agricultural sciences, energy, materials science, nanomedicine, and environmental science. The most effective strategy to revamp contemporary farming methods is the controlled synthesis of current nano-materials—a process that is simple, safe, and economically viable [1]. Precision agriculture is the latest innovation in modern farming, made possible by state-of-the-art nanomaterials that may be found in nature in plants and soil. Natural resource depletion, pest disease outbreaks, and changing weather patterns are only a few of the major threats to agricultural output [2,3,4].

Food and Agriculture Organization projections put the global population at 9–10 billion by 2050, meaning food production has to increase by 25–70% from where it is now [5]. Therefore, new technology must be used in the agricultural sector to guarantee sustainability and production in order to feed the growing population. By introducing a nano-based smart delivery system that revamps agriculture and associated industries, nanotechnology might play a role in the new technology-based agricultural revolution [3,6,7,8]. According to what is known, NPs of different sizes, shapes, and kinds may improve stance varieties, pesticide Nano formulation, plant disease diagnostics, and more [9].

A plethora of NPs with ever-improving capabilities and applications are unveiled

annually. The biological responses to NPs are determined by their physicochemical features, which include their size, zeta potential, and concentration [10,11]. The plant productivity could be improved with the help of NPs because of their many potential uses, including as germination enhancers, in the creation of nano fertilizers, as herbicide delivery systems, as nano sensors for pest detection, and as nanoporous zeolites for slow release and efficient water and fertilizer dosage [12,13]. However, some NPs exhibit phytotoxic effects, meaning they hinder seed germination or are toxic to seedlings [14,15,16].

Leaching, hydrolysis, degradation by photolysis, and decomposition make certain fertilizers inaccessible to plants, despite the fact that they are an essential source for plant growth and development. Nano pesticides and nano fertilizers are only two examples of the many novel NP solutions developed in recent years with the goal of lowering food waste and raising crop yields [2,6,17].

Nano fertilizers and nano encapsulated nutrients control the release of chemical fertilizers that enhance the target plant activity [18,19]. Multiple NPs are being evaluated for their ability to protect plants from different environmental stresses and to support plant growth [20]. In plant biotechnology, this field of study opens up new possibilities for influencing gene expression as well as cellular and cellular organelle properties. In addition to their many uses in agriculture and environmental remediation; NPs have a wide range of biosensor applications [21].

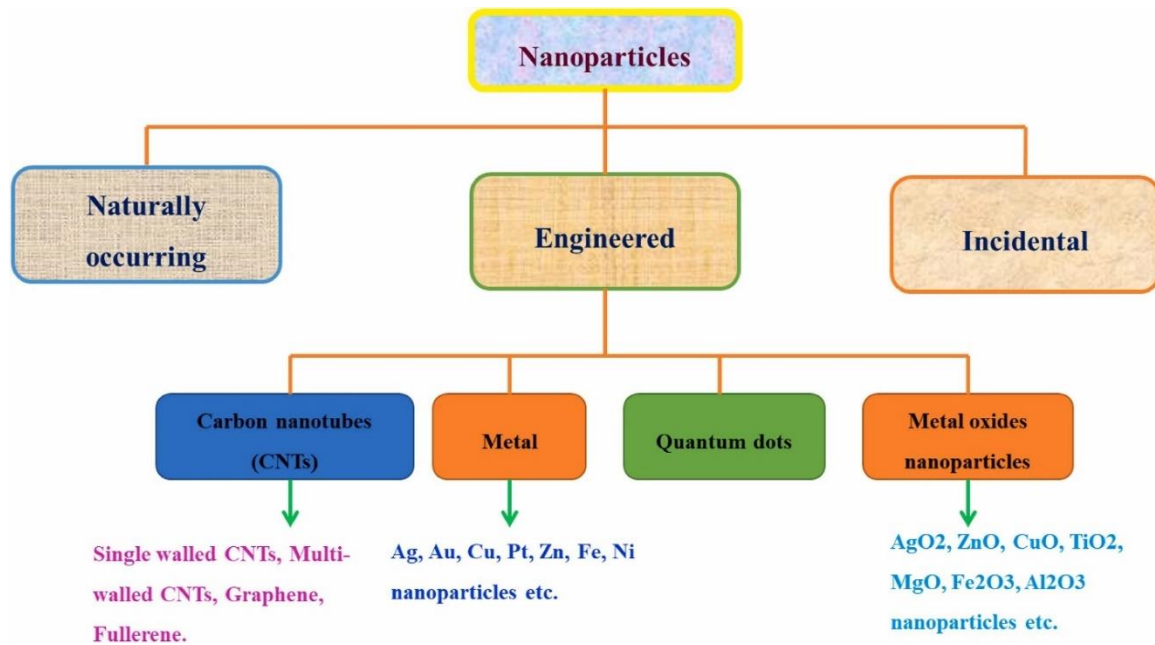


Fig. 1. Nanoparticles and their types

Despite their usefulness in agriculture, NPs have been shown to cause phytotoxicity and serious environmental problems. Anthropogenic activities release them into the environment, where they enter the food chain and produce biomagnification [22]. Nanoparticles (NPs) have a significant impact on plant uptake and translocation due to their size, concentration, types, toxicity, surface charge, pore sizes, reactivity, and other properties [23,24]. There is a potential for NPs to alter their characteristics, reactivity, and bioavailability to live organisms when they penetrate treated surfaces. The purpose of this review is to provide a balanced account of the pros and cons of using nanoscale materials in farming.

2. PLANT ABSORPTION OF NANOPARTICLES

In order to prevent the entrance of any foreign material, including NPs, the cell walls of plants include a variety of functional groups, including as carboxylate, hydroxyl, phosphate, and many more. These groups combine to form biomolecules, such as protein, polysaccharides, and cellulose [25]. The plant species is the primary determinant of NP uptake and translocation. Therefore, NPs enter plants by a process involving the whole system, including roots, stems, and leaves, which interact with soil, water, and other environmental variables. Additionally, NPs in soil may cause root system

interactions that result in cellular absorption [26,27]. Only NPs with a diameter similar to that of the cell wall may pass through its sieving capabilities and reach the plasma membrane. The cell wall diameter ranges from 5 to 20 nm. Following a complicated chain reaction, the NPs cross the root cell membrane, enter the plant's vascular system, and eventually make their way to the leaves [28,29]. Nanoparticles of a certain size may diffuse across lipid bilayers and enter cells by endocytosis via pore creation, binding to ion channels and aquaporins, and so on [30].

There are two pathways that NPs may take after they enter a plant cell: the apoplastic and the symplastic transport systems (Fig. 2). The size of the pore determines how NPs enter the cell wall; hence, smaller NPs move more freely [31], but bigger particles pass via stomata, hydathodes, and the stigma of the flower [32]. Although there are many stomata that can open and close, only a small fraction of them really can. Nanoparticles (NPs) larger than 40 nm are able to cross the plant's stomata and hydathodes on their way to the surface of the leaf, where they translocate via the leaf phloem and palisade parenchyma [33]. The seed coat has parenchymatous intercellular gaps that the NPs may penetrate [34]. On the other hand, aquaporins have a role in controlling NP entrance in the seed coat by reassembling the AQP-1 and Galphai-3 regulatory complex [35].

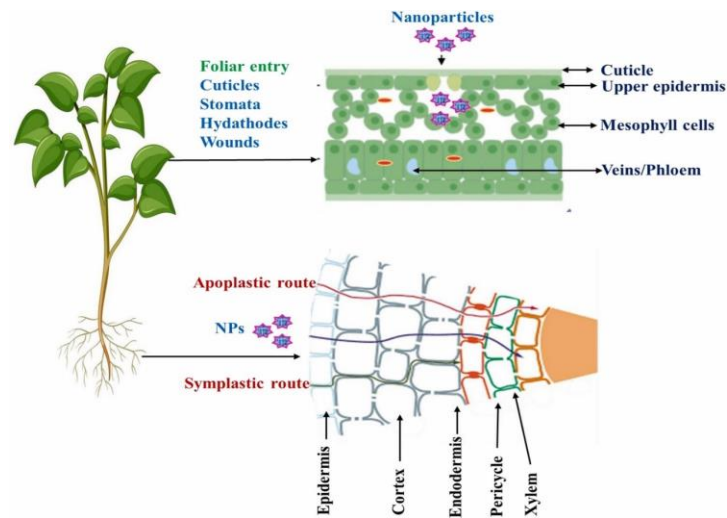


Fig. 2. An organized display of NPs absorption and translocation into various plant tissues via apoplastic and symplastic pathways, respectively, according to the plant's entrance point (leaves and roots)

3. NANOPARTICLES INFLUENCE ON PLANTS

Fig. 3 explains how NPs change plant shape by interfering with plant metabolism via several pathways, providing micronutrients, and regulating genes. Many different types of pathogens may infect crops, causing illnesses and reducing crop yields and economic output. The non-phytotoxicity, wide availability, and low cost of NPs make them useful in many agricultural contexts. Numerous plant species benefit from the application of NPs at pre-optimized rates, which enhance seed germination, stand establishment,

growth, and yield production. Plants build defense mechanisms by regulating molecular, biochemical, and physiological pathways in response to the many stresses they encounter during their life cycle. Plants address these challenges by adjusting gene expression in specific ways, which they call molecular pathways. Table 1 shows the results of many research that show the impact of NPs on plant growth and development is concentration dependent. NPs increase the activity of antioxidant enzymes such as peroxidase (POD), superoxide dismutase (SOD), and catalase (CAT) [36].

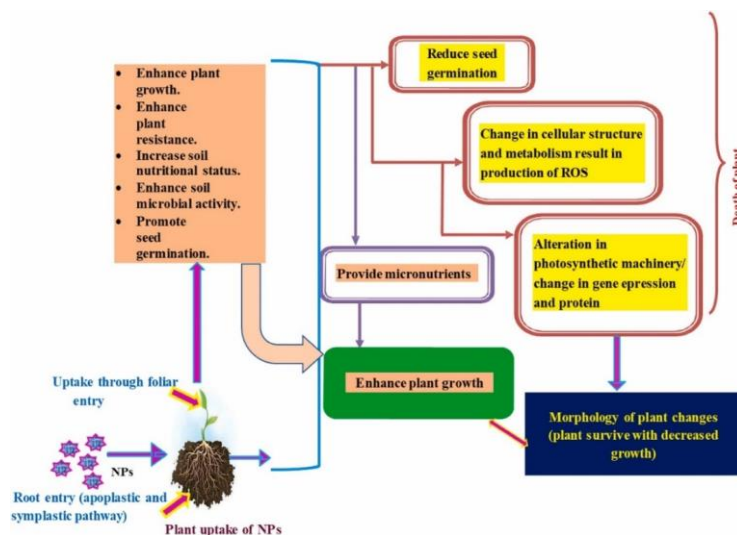


Fig. 3. NPs may interact with plant metabolism in a variety of ways or interfere with different plant oxidative processes

4. IMPACT OF DIFFERENT NPS ON THE PHYSIOLOGICAL PROCESSES INVOLVED IN PLANT DEVELOPMENT, GROWTH, AND MATURATION

4.1 Copper Nanoparticles

Various plant species were shown to have their germination, biomass, shoot development, and other processes significantly impacted by CuO NP exposure [37,38,39]. There was no effect on seed germination caused by CuO NP toxicity in maize plants. The NPs were translocated to the shoots by xylem and then returned to the roots via phloem [40]. According to [41], seedlings of *B. napus* were grown in MS media with CuO NPs for 10 days. The highest dosage of 10 mg L⁻¹ resulted in an induction of growth, whereas higher concentrations (100 and 1000 mg L⁻¹) resulted in a reduction in root dry weight and shoot elongation.

Culture medium hindered the development of *Lemna minor* at lower concentrations compared to higher concentrations of CuO [42]. The effect on the *Mentha longifolia* plant was a 45-48.4% rise in height and growth, a 29.4-33.9% increase in internodes, a 55.6-26.2% increase in shoots, and a 30-40% increase in reproduction coefficient when a colloidal solution of CuNP (0.5 mg L⁻¹) and CoNP (0.8 mg L⁻¹) and MS media were applied to the plant [43]. Copper nanoparticles, which are biosynthesized from tea extract, had positive impacts on the development of seedlings and nitric oxide signalling when exposed to *Lactuca sativa* at a concentration of 20 µg mL⁻¹ or less [44].

4.2 Iron Oxide Nanoparticles

When applied to plants, iron oxide, NPs significantly improve their development, stress tolerance, and nutritional status. In a study conducted by [45], it was found that soaking wheat (*T. aestivum*) in distilled water and then incubating it in a solution containing iron nanoparticles improved the germination percentage. However, when the roots were soaked in distilled water without the NPs, root growth was reduced, but when the roots were soaked in distilled water with the NPs suspension, root growth was enhanced. At concentrations of 3 and 25 the physiology of *A. thaliana* was impacted by both the positive and negative charged ions of iron oxide. The seedling and root length were unaffected by a dose of 3 mg. But they were severely decreased at 25 mg [46].

Iron oxide and chelated iron EDTA treatments of *Acinetobacter hypogaea* increase peanut plant biomass, germination, and growth via increasing enzyme antioxidant activities and phytohormone levels. According to [47], applying to plants increased their availability of iron, and the author even advised using it as a fertilizer. Root elongation in *L. sativa* seedlings was found to be improved by 12-26% when exposed to Fe₂O₃ NP (5-20 ppm), as reported by [48]. A study conducted that when Fe₃O₄ was accumulated in *Hordeum vulgare*, plant growth and photosynthetic efficiency were both improved [49].

4.3 Silver Nanoparticles

The antibacterial properties of silver (Ag) have led to its increased exposure to both plants and people as a result of its widespread usage in industry and medicine. Ag NPs have many beneficial effects on plant growth and development, and their use in agriculture has shown encouraging results [50]. Overuse of silver nanoparticles boosted the production and activity of antioxidants such as proline and carotenoids, as well as peroxidases and catalases. Also, it decreased the root length of *V. radiata* and *Sorghum bicolor* and improved seed germination and development in *Lolium multiflorum* and *Eruca sativa* at higher doses [52,53]. Separately, [54] revealed that Ag NPs alleviated heat stress symptoms in *T. aestivum*. Plants treated with Ag NPs had improvements in many biochemical parameters, including leaf area, root and shoot length, carbohydrate and protein contents, and activity of antioxidant enzymes. These plants were *B. juncea*, maize, and common bean [55].

4.4 Carbon Nanotubes

The diagnostic, biomedical, and agricultural communities are showing increasing interest in carbon nanotubes (CNTs) due to their diverse physicochemical characteristics. Carbon nanotubes (CNTs) have unique physicochemical characteristics that make them excellent plant growth regulators, water-absorbing agents, and nutritional supplementers [56]. In light of this, there are a number of scientific applications for carbon-containing NPs, SWCNTs enhance water intake and accelerate germination rate in rice seedlings via modulating gene expression [57].

CNTs, in comparison to a control group, increased the rate of germination and development of tomato seedlings (*Solanum*

lycopersicum) and helped them absorb water via piercing their protective outer layer [58]. *B. juncea* may be effectively treated with multi-walled carbon nanotubes (~30 nm) using a reduced concentration of oxidized MWCNT [59]. MWCNTs hasten the germination of seeds in *G. max* and *H. vulgare* without negatively impacting the plants' subsequent development [60]. Additionally, compared to control seeds, treated seeds showed an increase in genes encoding water channel protein. Water delivery in *Z. mays* plants was enhanced by using a lower concentration of MWCNTs [61].

According to other researchers [59,62], the same outcomes have been shown for *B. napus* and *C. arietinum* plants. In addition, under NaCl challenged circumstances, MWCNTs were shown to improve aquaporin transduction by altering the lipid content, stiffness, and permeability of the root plasma membrane [63].

5. THE STRUCTURE AND FUNCTION OF PLANTS' PHOTOSYNTHETIC SYSTEM ARE AFFECTED BY NANOPARTICLES

The process of converting solar energy into chemical energy is carried out by both plants and

algae. Just 2-4% of the energy is transformed by plants throughout their life cycle [64]. Plants play a crucial role in the oxygen cycle, translocate minerals and other vital nutrients to other parts of the food web, and carry out photosynthetic activities. Plants may take in both necessary and non-essential nutrients, but there is a threshold concentration beyond which they become poisonous [65]. Scientists are able to enhance plants' photosynthetic apparatus and efficiency via gene editing and the use of nanotechnology. Translocation of NPs and the acceleration of plant biotechnology are both impacted by the inevitable interaction between plants and NPs.

Toxic NPs, such as CuO and Ag, disrupt photosynthetic apparatus structure and function. Reduced photosynthetic pigment concentration (especially chlorophyll), grana disruption, and other chloroplast abnormalities are all effects of the NPs. Photosynthesis and photosystem II are both made less efficient by NPs. Despite the fact that NPs of CeO₂ and TiO₂ enhanced electron transport between PS II and I and Rubisco activity, they did not eliminate all negative effects [66]. The use of SWCNTs tripled the

Table 1. The impact of several NPs on the photosynthetic apparatus organization

NPs	Plants Species	Concentration	Effect	References
CuO	<i>Lemna gibba</i>	1.1–0.4 g L ⁻¹	Photosynthetic pigment reduces.	[69]
	<i>Elodea densa</i>	1 mg L ⁻¹	Broken thylakoid membrane and chloroplast water oxidizing complex.	[70]
	<i>Elsholtzia splendens</i>	100 mg L ⁻¹	Fewer photosynthetic pigments.	[71]
	<i>Oryza sativa</i>	10 mg L ⁻¹	Reduced photosynthetic pigments and thylakoid quantity per granum.	[72]
Ag, NP	<i>Chlamydomonas reinhardtii</i>	2 μM	Reduced electron transit and increased QB non-reducing centers.	[73]
	<i>Skeletonema costatum</i>	5 mg L ⁻¹	Photosynthesis inhibition and chl a reduction.	[74]
	<i>Spirodela polyrhiza</i>	25 mg L ⁻¹	Plastoquinone and chl a fluorescence decrease.	[75]
	<i>Wolffia globosa</i>	10 mg L ⁻¹	Reductions in chlorophyll a (chl a) of 77.7 percent, carotenoids of 66.2 percent, and soluble proteins of 72.9 percent were observed.	[76]
TiO ₂	<i>Spinacia oleracea</i>	0.25%	Total chlorophyll increased.	[77]
	<i>Chlorella</i> sp	1 mg L ⁻¹	Less chlorophyll and changes to the chloroplast, plasma membrane, and nucleus.	[78]
CeO ₂	<i>Zea mays</i>	400 mg kg ⁻¹	Chlorophyll a content become reduced	[79]
	<i>Solanum lycopersicum</i>	250 mg kg ⁻¹	Chlorophyll a and b content increase.	[80]
	<i>Phaseolus vulgaris</i>	250 mg kg ⁻¹	Decrease content of chlorophyll and carotenoid.	[81]
ZnS	<i>Brassica juncea</i>	25 mg kg ⁻¹	Decline the presence of Chlorophyll a and b content.	[82]

photosynthetic activity and electron transport rate in chloroplasts. Increased photosynthetic carbon absorption is facilitated by nano TiO₂ induced carboxylation via Rubisco activation [66]. SiO₂ NPs accelerated photosynthesis via changing the activity of carbonic anhydrase and photosynthetic pigments [67,68].

6. NANOPARTICLES AND DEFENSE MECHANISM

It has been shown that NP exposure may trigger oxidative damage, ROS generation, and antioxidant defense system activation [83]. Enzymatic antioxidants like glutathione reductase (GR), glutathione, ascorbate, thiols, and phenolics are part of the antioxidant defense, along with enzymatic antioxidants like APOX, CAT, SOD, GPOX, and GR [83,84,85]. Superoxide dismutase (SOD) catalyses the conversion of superoxide ions into hydrogen peroxide, whereas peroxy radicals and reactive oxygen species (ROS) are stifled by CAT and GPOX, respectively [83]. Direct reduction of H₂O₂ into H₂O occurs during the formation of ROS by NPs via APOX [83,86]. To combat the oxidative stress caused by NPs, [Wei and Wang (2013)] examined the plants that showed promise as anti-oxidants. The anti-oxidant enzyme capabilities of several NPs have been studied by [87]. For example, nFe₂O₄, nCeO₂, and nCo₃O₄ stimulate catalase, nFe₃O₄, nCeO₂, nMnO₂, nCuO, and nAu promote GPOX, and nCeO₂ and fullerene produce SOD.

Although many nano phytotoxicity studies have shown that plants exposed to NPs have enzyme activity disturbances, no evidence has been found to link these disturbances to the chemical properties of NPs or to prove that the enzyme interactions with the NPs were the cause of these changes. Indeed, research revealed that NPs had varying impacts on enzyme activity. While nTiO₂ increased the activities of GPOX, SOD, and CAT in Lemna minor [88] and SOD, CAT, APOX, and CAT in spinach [89], it lowered the activities of GR and APOX in Vicia faba [90]. Because of this, it is not easy to determine which NPs have an effect on particular enzymes.

7. CONCLUSION

Nanotechnology is a relatively new method that has found applications in several scientific disciplines. Nanoparticles (NPs) are a potential new material for food security and cutting-edge farming techniques. Incorporating NPs into

farming practices boosts the world economy in several ways. Toxic effects of NPs are not yet understood because of a lack of adequate information, while NPs-plant interaction is sensitive to NP size and may have both beneficial and detrimental effects. Their effects may change depending on the plant's development stage, exposure duration, uptake rate, and physiochemical characteristics. In comparison to more conventional resources, the advent of NPs has increased efficacy and agronomic efficiency. When it comes to detecting diseases on-site, the interactions between plants and NPs provide genuine promise for achieving sustainable agriculture. There has been research into the potential of nano-based formulations, including as herbicides, insecticides, fertilizers, fungicides, and sensors, for improved plant management and controlled release to safeguard the environment. However, environmental contamination is a major worry due to the growing usage of NPs in agriculture and related industries, thus proactive steps should be made to prevent their accumulation. Food security is a major concern for agricultural scientists due to the increasing human population. Not only will the nano revolution help with food security and environmental preservation, but it is also predicted to bring about a paradigm change in the sustainability of agriculture. To lessen the phytotoxic effects and increase agricultural output for human welfare, molecular science research into the interactions between plants and NPs is urgently required.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Abu-Hamdah R, Cho WJ, Cho SJ, Jeremic A, Kelly M, Ilie AE, Jena BP. Regulation of the water channel aquaporin-1: Isolation and reconstitution of the regulatory complex. *Cell Biology International*. 2004; 28(1):7-17.
2. Acharya P, Jayaprakasha GK, Crosby KM, Jifon JL, Patil BS. Green-synthesized nanoparticles enhanced seedling growth, yield, and quality of onion (*Allium cepa* L.) *ACS Sustain. Chem. Eng.* 2020;7:14580-14590.
3. Adhikari T, Kundu S, Biswas AK, Tarafdar JC, Rao AS. Effect of copper oxide nano particle on seed germination of selected

- crops. Journal of Agricultural Science and Technology. A. 2012;2(6A):815.
4. Almutairi ZM. Effect of nano-silicon application on the expression of salt tolerance genes in germinating tomato (*Solanum lycopersicum* L.) seedlings under salt stress. Plant Omics. 2016; 9(1):106-114.
 5. Barrios AC, Rico CM, Trujillo-Reyes J, Medina-Velo IA, Peralta-Videa JR, Gardea-Torresdey JL. Effects of uncoated and citric acid coated cerium oxide nanoparticles, bulk cerium oxide, cerium acetate, and citric acid on tomato plants. Science of the total environment. 2016;563:956-964.
 6. Batsmanova LM, Gonchar LM, Taran NY, Okanenko AA. Using a colloidal solution of metal nanoparticles as micronutrient fertilizer for cereals Proceedings of the International Conference on Nanomaterials: Applications and Properties. Crimea, Ukraine. 2013;16-21.
 7. Bombin S, LeFebvre M, Sherwood J, Xu Y, Bao Y, Ramonell KM. Developmental and reproductive effects of iron oxide nanoparticles in Arabidopsis thaliana International Journal of Molecular Sciences. 2015;24174-24193.
 8. Camara MC, Campos EVR, Monteiro RA, do Espirito Santo Pereira A, de Freitas Proença, PL, & Fraceto LF. Development of stimuli-responsive nano-based pesticides: Emerging opportunities for agriculture. Journal of nanobiotechnology. 2019; 17(1):1-19.
 9. Da Costa MVJ, Sharma PK. Effect of copper oxide nanoparticles on growth, morphology, photosynthesis, and antioxidant response in *Oryza sativa*. Photosynthetica. 2016;54:110-119.
 10. Das A, Das B. Nanotechnology a potential tool to mitigate abiotic stress in crop plants. Abiotic and biotic stress in plants, Alexandre Bosco de Oliveira, IntechOpen; 2019. DOI: 10.5772/intechopen. 83562.
 11. De La Torre-Roche, R., Cantu, J., Tamez, C., Zuverza-Mena, N., Hamdi, H., Adisa, I. O., ... & White JC. Seed biofortification by engineered nanomaterials: A pathway to alleviate malnutrition? Journal of Agricultural and Food Chemistry. 2020;68(44):12189-12202.
 12. DeRosa MC, Monreal, C, Schnitzer M, Walsh R, Sultan Y. Nanotechnology in fertilizers. Nature nanotechnology. 2010; 5(2):91-91.
 13. Falco WF, Scherer MD, Oliveira SL, Wender H, Colbeck I, Lawson T, Caires AR. Phytotoxicity of silver nanoparticles on *Vicia faba*: evaluation of particle size effects on photosynthetic performance and leaf gas exchange. Science of The Total Environment. 2020;701:134816.
 14. Fleischer A, O'Neill MA, Ehwald R. The pore size of non-graminaceous plant cell walls is rapidly decreased by borate ester cross-linking of the pectic polysaccharide rhamnogalacturonan II. Plant Physiology. 1999;121(3):829-838.
 15. Foltête AS, Masfarau JF, Bigorgne E, Nahmani J, Chaurand P, Botta C, Cotelle S. Environmental impact of sunscreen nanomaterials: ecotoxicity and genotoxicity of altered TiO₂ nanocomposites on *Vicia faba*. Environmental pollution. 2011; 159(10):2515-2522.
 16. Gao F, Hong F, Liu C, Zheng L, Su M, Wu X, Yang P. Mechanism of nano-anatase TiO₂ on promoting photosynthetic carbon reaction of spinach: Inducing complex of rubisco-rubisco activase. Biological trace element research. 2006;111:239-253.
 17. Hao Y, Zhang Z, Rui Y, Ren JY, Hou TQ, Wu SJ, Liu LM. Effect of different nanoparticles on seed germination and seedling growth in rice Advance Engineering Research. 2016;85:166-173.
 18. Hayes KL, Mui J, Song B, Sani ES, Eisenman SW, Sheffield JB, Kim B. Effects, uptake, and translocation of aluminum oxide nanoparticles in lettuce: A comparison study to phytotoxic aluminum ions. Science of the Total Environment. 2020;719:137393.
 19. Hossain Z, Mustafa G, Sakata K, Komatsu S. Insights into the proteomic response of soybean towards Al₂O₃, ZnO, and Ag nanoparticles stress. Journal of hazardous materials. 2016;304:291-305.
 20. Hu P, An J, Faulkner MM, Wu H, Li Z, Tian X, Giraldo JP. Nanoparticle charge and size control foliar delivery efficiency to plant cells and organelles. ACS nano. 2020;14(7):7970-7986.
 21. Huang J, Cheng J, Yi J. Impact of silver nanoparticles on marine diatom *Skeletonema costatum*. Journal of Applied Toxicology. 2016;36(10):1343-1354.
 22. Iqbal M, Raja NI, Hussain M, Ejaz M, Yasmeen F. Effect of silver nanoparticles on growth of wheat under heat stress Iranian Journal of Science & Technology: Sci. 2019;43(2):387-395.

23. Iswarya V, Bhuvaneshwari M, Alex SA, Iyer S, Chaudhuri G, Chandrasekaran PT, Mukherjee A. Combined toxicity of two crystalline phases (anatase and rutile) of Titania nanoparticles towards freshwater microalgae: *Chlorella* sp. Aquatic toxicology. 2015;161:154-169.
24. Kah M, Tufenkji N, White JC. Nano-enabled strategies to enhance crop nutrition and protection. Nature nanotechnology. 2019;14(6):532-540.
25. Kaur N, Kaur J, Grewal SK, Singh I. Effect of heat stress on antioxidative defense system and its amelioration by heat acclimation and salicylic acid pre-treatments in three pigeonpea genotypes. Indian Journal of Agricultural Biochemistry. 2019;32(1):106-110.
26. Ke W, Xiong ZT, Chen S, Chen J. Effects of copper and mineral nutrition on growth, copper accumulation and mineral element uptake in two *Rumex japonicus* populations from a copper mine and an uncontaminated field site. Environmental and Experimental Botany. 2007;59(1):59-67.
27. Khan I, Saeed K, Khan I. Nanoparticles: Properties, applications and toxicities. Arabian journal of chemistry. 2019; 12(7):908-931.
28. Khodakovskaya M, Dervishi E, Mahmood M, Xu Y, Li Z, Watanabe F, Biris AS. Carbon nanotubes are able to penetrate plant seed coat and dramatically affect seed germination and plant growth. ACS nano. 2009;3(10):3221-3227.
29. Kirschbaum MU. Does enhanced photosynthesis enhance growth? Lessons learned from CO₂ enrichment studies. Plant physiology. 2011;155(1):117-124.
30. Krishnaraj C, Ramachandran R, Mohan K, Kalaichelvan PT. Optimization for rapid synthesis of silver nanoparticles and its effect on phytopathogenic fungi. Spectrochemical Acta Part A: Molecular and Biomolecular Spectroscopy. 2012; 93:95-99.
31. Lahiani MH, Dervishi E, Chen J, Nima Z, Gaume A, Biris AS, Khodakovskaya MV. Impact of carbon nanotube exposure to seeds of valuable crops. ACS applied materials & interfaces. 2013;5(16):7965-7973.
32. Lee CW, Mahendra S, Zodrow K, Li D, Tsai YC, Braam J, Alvarez PJ. Developmental phytotoxicity of metal oxide nanoparticles to *Arabidopsis thaliana*. Environmental Toxicology and Chemistry: An International Journal. 2010;29(3):669-675.
33. Lee WM, An YJ, Yoon H, Kweon HS. Toxicity and bioavailability of copper nanoparticles to the terrestrial plants mung bean (*Phaseolus radiatus*) and wheat (*Triticum aestivum*): Plant agar test for water-insoluble nanoparticles. Environmental Toxicology and Chemistry: An International Journal. 2008;27(9):1915-1921.
34. Lei Z, Mingyu S, Xiao W, Chao L, Chunxiang Q, Liang C, Fashui H. Antioxidant stress is promoted by nano-anatase in spinach chloroplasts under UV-B radiation. Biological Trace Element Research. 2008;121:69-79.
35. Liu R, Zhang H, Lal R. Effects of stabilized nanoparticles of copper, zinc, manganese, and iron oxides in low concentrations on lettuce (*Lactuca sativa*) seed germination: Nanotoxicants or nanonutrients. Water, Air, & Soil Pollution. 2016;227:1-14.
36. Lowry GV, Avellan A, Gilbertson LM. Opportunities and challenges for nanotechnology in the agri-tech revolution. Nature nanotechnology. 2019;14(6):517-522.
37. Majumdar S, Peralta-Videa JR, Trujillo-Reyes J, Sun Y, Barrios AC, Niu G, Gardea-Torresdey JL. Soil organic matter influences cerium translocation and physiological processes in kidney bean plants exposed to cerium oxide nanoparticles. Science of the Total Environment. 2016;569:201-211.
38. Martinez-Ballesta MC, Chelbi N, Lopez-Zaplana A, Carvajal M. Discerning the mechanism of the multiwalled carbon nanotubes effect on root cell water and nutrient transport. Plant Physiology and Biochemistry. 2020;146:23-30.
39. Matorin DN, Todorenko DA, Seifullina NK, Zayadan BK, Rubin AB. Effect of silver nanoparticles on the parameters of chlorophyll fluorescence and P 700 reaction in the green alga *Chlamydomonas reinhardtii*. Microbiology. 2013;82:809-814.
40. Mittler R. ROS is good. Trends in plant science. 2017;22(1):1-19.
41. Morteza E, Moaveni P, Farahani HA, Kiyani M. Study of photosynthetic pigments changes of maize (*Zea mays* L.) under nano TiO₂ spraying at various growth stages. Springer Plus. 2013;2:1-5.
42. Nair R, Varghese SH, Nair BG, Maekawa T, Yoshida Y, Kumar DS. Nanoparticulate

- material delivery to plants. Plant science. 2010;179(3):154-163.
43. Nayan R, Rawat M, Negi B, Pande A, Arora S. Zinc sulfide nanoparticle mediated alterations in growth and antioxidant status of Brassica juncea. Biologia. 2016;71(8):896-902.
 44. Nekrasova GF, Ushakova OS, Ermakov AE, Uimin MA, Byzov IV. Effects of copper (II) ions and copper oxide nanoparticles on Elodea densa Planch. Russian Journal of Ecology. 2011;42:458-463.
 45. Palocci C, Valletta A, Chronopoulou L, Donati L, Bramosanti M, Brasili E, Pasqua G. Endocytic pathways involved in PLGA nanoparticle uptake by grapevine cells and role of cell wall and membrane in size selection. Plant cell reports. 2017;36:1917-1928.
 46. Pelegrino MT, Kohatsu MY, Seabra AB, Monteiro LR, Gomes DG, Oliveira HC, Lange CN. Effects of copper oxide nanoparticles on growth of lettuce (*Lactuca sativa* L.) seedlings and possible implications of nitric oxide in their antioxidative defense. Environmental Monitoring and Assessment. 2020;192:1-14.
 47. Pérez-de-Luque A. Interaction of nanomaterials with plants: What do we need for real applications in agriculture. Frontiers in Environmental Science. 2017;5:12.
 48. Perreault F, Oukarroum A, Pirastru L, Sirois L, Matias WG, Popovic R. Evaluation of copper oxide nanoparticles toxicity using chlorophyll a fluorescence imaging in *Lemna gibba*. Journal of Botany; 2010.
 49. Rai PK, Kumar V, Lee S, Raza N, Kim KH, Ok YS, Tsang DC. Nanoparticle-plant interaction: Implications in energy, environment, and agriculture. Environment international. 2018;119:1-19.
 50. Rajput VD, Minkina T, Kumari A, Harish Singh VK, Verma KK, Keswani C. Coping with the challenges of abiotic stress in plants: New dimensions in the field application of nanoparticles. Plants. 2021;10(6):1221.
 51. Rajput VD, Minkina T, Suskova S, Mandzhieva S, Tsitsuashvili V, Chapligin V, Fedorenko A. Effects of copper nanoparticles (CuO NPs) on crop plants: A mini review. Bio Nanoscience. 2018;8:36-42.
 52. Rajput V, Minkina T, Fedorenko A, Sushkova S, Mandzhieva S, Lysenko V, Ghazaryan K. Toxicity of copper oxide nanoparticles on spring barley (*Hordeum sativum* distichum). Science of the Total Environment. 2018;645:1103-1113.
 53. Rico CM, Majumdar S, Duarte-Gardea M, Peralta-Videa JR, Gardea-Torresdey JL. Interaction of nanoparticles with edible plants and their possible implications in the food chain. Journal of agricultural and food chemistry. 2011;59(8):3485-3498.
 54. Rico CM, Peralta-Videa JR, Gardea-Torresdey JL. Chemistry, biochemistry of nanoparticles, and their role in antioxidant defense system in plants. Nanotechnology and plant sciences: nanoparticles and their impact on plants. 2015;1-17.
 55. Rizwan M, Ali S, Qayyum MF, Ok YS, Adrees M, Ibrahim M, Abbas F. Effect of metal and metal oxide nanoparticles on growth and physiology of globally important food crops: A critical review. Journal of hazardous materials. 2017;322:2-16.
 56. Salama HM. Effects of silver nanoparticles in some crop plants, common bean (*Phaseolus vulgaris* L.) and corn (*Zea mays* L.). Int Res J Biotechnol. 2012;3(10):190-197.
 57. Schmidt J. Nanoparticle-induced membrane pore formation studied with lipid bilayer arrays. Biophysical Journal. 2015;108(2):344a-345a.
 58. Scott NR. Nanotechnology opportunities in agriculture and food systems. In Biological & Environmental Engineering, Cornell University NSF Nanoscale Science & Engineering Grantees Conference. 2007;5.
 59. Scott NR, Chen H, Cui H. Nanotechnology applications and implications of agrochemicals toward sustainable agriculture and food systems. Journal of agricultural and food chemistry. 2018;66(26):6451-6456.
 60. Scrinis G, Lyons K. The emerging nanocorporate paradigm: Nanotechnology and the transformation of nature, food and agri-food systems. The International Journal of Sociology of Agriculture and Food. 2007;15(2):22-44.
 61. Shabnam N, Sharmila P, Pardha-Saradhi P. Impact of ionic and nanoparticle speciation states of silver on light harnessing photosynthetic events in *Spirodela polyrhiza*. International Journal of Phytoremediation. 2017;19(1):80-86.

62. Shakiba S, Astete CE, Paudel S, Sabliov CM, Rodrigues DF, Louie SM. Emerging investigator series: polymeric nanocarriers for agricultural applications: synthesis, characterization, and environmental and biological interactions. *Environmental Science: Nano*. 2020;7(1):37-67.
63. Shi J, Peng C, Yang Y, Yang J, Zhang H, Yuan X, Hu T. Phytotoxicity and accumulation of copper oxide nanoparticles to the Cu-tolerant plant *Elsholtzia splendens*. *Nanotoxicology*. 2014;8(2):179-188.
64. Siddiqui MH, Al-Wahaibi MH. Role of nano-SiO₂ in germination of tomato (*Lycopersicon esculentum* seeds Mill.). *Saudi Journal of Biological Sciences*. 2014;21(1):13-17.
65. Singh A, Singh S, Prasad SM, Tripathi DK, Singh VP, Ahmad P, Chauhan DK, Prasad SM. Silicon and nanotechnology role in agriculture and future perspective in silicon in plants in advances and future prospects, CRC Press. 2016;392.
66. Singh J, Lee BK. Influence of nano-TiO₂ particles on the bioaccumulation of Cd in soybean plants (*Glycine max*): A possible mechanism for the removal of Cd from the contaminated soil. *Journal of environmental management*. 2016;170:88-96.
67. Talankova-Sereda TE, Liapina KV, Shkopinskij EA, Ustinov AI, Kovalyova AV, Dulnev PG, Kucenko NI. The influence of Cu and Co nanoparticles on growth characteristics and biochemical structure of *Mentha longifolia* in vitro *Nanosci. Nanoeng.* 2016;4:31-39.
68. Tighe-Neira R, Carmora E, Recio G, Nunes-Nesi A, Reyes -Diaz M, Alberdi M, Inostroza-Blancheteau C. Metallic nanoparticles influence the structure and function of the photosynthetic apparatus in plants *Plant Physiology & Biochemistry*. 2018;130:408-417.
69. Tiwari DK, Dasgupta-Schubert N, Villaseñor Cendejas LM, Villegas J, Carreto Montoya L, Borjas García SE. Interfacing carbon nanotubes (CNT) with plants: enhancement of growth, water and ionic nutrient uptake in maize (*Zea mays*) and implications for nanoagriculture. *Appl. Nanosci.* 2014;4(5):577-591.
70. Tombuloglu H, Slimani Y, Tombuloglu G, Almessiere M, Baykal A. Uptake and translocation of magnetite (Fe₃O₄) nanoparticles and its impact on photosynthetic genes in barley (*Hordeum vulgare* L.). *Chemosphere*. 2019;226:110-122.
71. Tripathi A, Liu S, Singh PK, Kumar N, Pandey AC, Tripathi DK, Sahi S. Differential phytotoxic responses of silver nitrate (AgNO₃) and silver nanoparticle (AgNps) in *Cucumis sativus* L. *Plant Gene*. 2017;11:255-264.
72. Tripathi DK, Mishra RK, Singh S, Singh S, Vishwakarma K, Sharma S, Chauhan DK. Nitric oxide ameliorates zinc oxide nanoparticles phytotoxicity in wheat seedlings: implication of the ascorbate–glutathione cycle. *Frontiers in plant science*. 2017;8:1.
73. Tripathi DK, Singh S, Singh S, Pandey R, Singh VP, Sharma NC, Chauhan DK. An overview on manufactured nanoparticles in plants: Uptake, translocation, accumulation and phytotoxicity. *Plant physiology and biochemistry*. 2017;110:2-12.
74. Tripathi S, Sonkar SK, Sarkar S. Growth stimulation of gram (*Cicer arietinum*) plant by water soluble carbon nanotubes. *Nanoscale*. 2011;3(3):1176-1181.
75. Vannini C, Domingo G, Onelli E, Prinsi B, Marsoni M, Espen L, Bracale M. Morphological and proteomic responses of *Eruca sativa* exposed to silver nanoparticles or silver nitrate. *PloS one*. 2013;8(7):e68752.
76. Vinopal S, Ruml T, Kotrba P. Biosorption of Cd²⁺ and Zn²⁺ by cell surface-engineered *Saccharomyces cerevisiae*. *International Biodeterioration & Biodegradation*. 2007;60(2):96-102.
77. Wang Z, Xie X, Zhao J, Liu X, Feng W, White JC, Xing B. Xylem-and phloem-based transport of CuO nanoparticles in maize (*Zea mays* L.). *Environmental science & technology*. 2012;46(8):4434-4441.
78. Wei H, Wang E. Nanomaterials with enzyme-like characteristics (nanozymes): Next-generation artificial enzymes. *Chemical Society Reviews*. 2013;42(14):6060-6093.
79. Xie Y, Li B, Zhang Q, Zhang C. Effects of nano-silicon dioxide on photosynthetic fluorescence characteristics of *Indocalamus barbatus* McClure J. *Nanjing For. Univ. (Nat. Sci. Ed.)*. 2012;2:59-63.
80. Yang F, Liu C, Gao F, Su M, Wu X, Zheng L, Yang P. The improvement of spinach growth by nano-anatase TiO₂ treatment is related to nitrogen photoreduction.

- Biological trace element research. 2007;119:77-88.
81. Yasmeen F, Razzaq A, Iqbal MN, Jhanzab HM. Effect of silver, copper and iron nanoparticles on wheat germination. *Int. J. Biosci.* 2015;6(4):112-117.
 82. Zhao L, Sun Y, Hernandez-Viezcas JA, Hong J, Majumdar S, Niu G, Gardea-Torresdey JL. Monitoring the environmental effects of CeO₂ and ZnO nanoparticles through the life cycle of corn (*Zea mays*) plants and in situ μ -XRF mapping of nutrients in kernels. *Environmental science & technology.* 2015;49(5):2921-2928.
 83. Zou X, Li P, Huang Q, Zhang H. The different response mechanisms of *Wolffia globosa*: Light-induced silver nanoparticle toxicity. *Aquatic Toxicology.* 2016;176: 97-105.
 84. Sakihama Y, Cohen MF, Grace SC, Yamasaki H. Plant phenolic antioxidant and prooxidant activities: phenolics-induced oxidative damage mediated by metals in plants. *Toxicology.* 2002 Aug 1;177(1):67-80.
 85. Alfieri ML, Panzella L, Amorati R, Cariola A, Valgimigli L, Napolitano A. Role of sulphur and heavier chalcogens on the antioxidant power and bioactivity of natural phenolic compounds. *Biomolecules.* 2022; 12(1):90.
 86. Zhang J, Ma J, Choksi TS, Zhou D, Han S, Liao YF, Yang HB, Liu D, Zeng Z, Liu W, Sun X. Strong metal-support interaction boosts activity, selectivity, and stability in electrosynthesis of H₂O₂. *Journal of the American Chemical Society.* 2022; 144(5):2255-63.
 87. Hanikoglu A, Ozben H, Hanikoglu F, Ozben T. Hybrid compounds & oxidative stress induced apoptosis in cancer therapy. *Current medicinal chemistry.* 2020; 27(13):2118-32.
 88. Vale G, Mehennaoui K, Cambier S, Libralato G, Jomini S, Domingos RF. Manufactured nanoparticles in the aquatic environment-biochemical responses on freshwater organisms: a critical overview. *Aquatic toxicology.* 2016;170: 162-74.
 89. Kumar V, Sharma M, Khare T, Wani SH. Impact of nanoparticles on oxidative stress and responsive antioxidative defense in plants. In *Nanomaterials in Plants, Algae, and Microorganisms.* Academic Press. 2018;393-406.
 90. Oufdou K, Benidire L, Lyubenova L, Daoui K, Fatemi ZE, Schröder P. Enzymes of the glutathione-ascorbate cycle in leaves and roots of rhizobia-inoculated faba bean plants (*Vicia faba* L.) under salinity stress. *European journal of soil biology.* 2014; 60:98-103.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/113997>