



Impact of Biofertilizers on Growth and Reproductive Performance of *Eisenia fetida* (Savigny 1926) During Flower Waste Vermicomposting Process

**D. Senthil Kumar^{1*}, P. Satheesh Kumar², V. Uthaya Kumar³,
and G. Anbuganapathi¹**

¹Department of Zoology, Kandaswami Kandar's College, Velur-638 182, Namakkal District, Tamil Nadu, India.

²Central Marine Fisheries Research Institute, Kochi-682018, Kerala, India.

³Department of Zoology, Bharathiyar University, Coimbatore, Tamil Nadu, India.

Authors' contributions

This work was carried out in collaboration between all authors. Author DSK designed the study, wrote the protocol, carried out the laboratory work and wrote the first draft of the manuscript. Authors PSK and VUK performed the statistical analysis and managed the literature searches. Author GA supervised the overall work. All authors read and approved the final manuscript

Research Article

Received 16th March 2013
Accepted 17th June 2013
Published 28th June 2013

ABSTRACT

The growth and reproductive performance of epigeic earthworm *Eisenia fetida* in biofertilizers enriched flower waste vermicomposting was studied for 80 days in the laboratory. Six feeding compositions – Co-Flower waste (FW) alone; A- FW + Earthworms (*E. fetida*); B- FW+ EW + Azospirillum (Azos); C- FW + EW+ Phosphobacteria (PB); D- FW+EW+ Blue green algae (BGA); E- FW + EW + Rhizobium (Rhizo). Maximum number of earthworms was recorded in C and minimum number in A, highest average body weight was observed in E and least in A. Maximum average body length was noted in B and minimum in A. Higher reproductive performance of earthworms under the influence of biofertilizers were clearly visible in C and E than A. The microbial populations and availability of food materials from biofertilizers enriched flower waste increases the biomass, average body weight, average body length and also increases the reproductive

*Corresponding author: Email: drskumar1968@gmail.com;

performance of earthworms in the vermicomposting process.

Keywords: *Eisenia fetida*; vermicompost; biofertilizers; reproductive performance.

ABBREVIATIONS

Azos - *Azospirillum*; PB – *Phosphobacteria*; BGA – *Blue green algae*; Rhizo – *Rhizobium*; Fw – *Flower waste*; TOC – *Total Organic Carbon*; TN – *Total Nitrogen*; VC – *Vermicomposting Coefficient*; Nrr – *Net reproductive rate*; RP – *Reproductive performance*.

1. INTRODUCTION

Every year approximately 40 MTY⁻¹ of organic waste produces worldwide. Disposal and environmental friendly management of these waste has become one of the serious global problems. Earthworms are one of the major soil macro invertebrates and are known for their contributions to soil formation and turn over with their widespread global distribution [1] Recycling of waste through vermicomposting, reduces the problem of non- utilization of live stock excreta [2]. Vermicomposting is one of the ways to reduce this organic waste and it has been adopted all over the world. Earthworms utilize the rich complex organic substances and to give humus like product [3]. There are several reports regarding the potential utilization of epigeic earthworms for successful degradation of organic wastes, generated from different industries such as paper and pulp industry [4,5] dairy industry [6]; winery and distillery industry [7]; wood and chips industry [8]; flower waste [9] and textile mill [10,11]. The results of these studies have shown that environmental conditions and earthworm population density affect earthworm growth and reproduction.

World-wide spread *E. fetida* was and still remains a favoured earthworm species for vermicomposting operations due to their wide range of tolerance for environmental variables. The growth patterns *E. fetida* in number of different organic waste resources have been investigated by various authors in laboratory culture [11,12]. *E. fetida* (Savigny) has been studied extensively, in particular for its potential in vermiculture [13]. Various aspects of the general biology and ecology of this species are well known now. The life cycle has been thoroughly investigated by [14], the moisture [15] and temperature requirements [16] are documented [17,18] and [19] also studied the reproduction of this species intensively.

In vermicomposting process the earthworms converting the organic wastes into valuable fertilizers and it is essential to know the influence of prevailing environmental factors on growth and reproduction of earthworms. The activities of earthworms are known to be influenced by quality of food [20] and moisture [21]. Among these food and environmental factors play an important role in the biology of earthworms. The moisture content of organic waste used in vermicomposting is an important parameter influencing the growth of the surface-feeding (epigeic) earthworm species *E. fetida*, since the earthworm's body contains about 80% water. This species gained weight maximally and survived best at temperatures between 20°C to 29°C and moisture content between 70% and 85% in horse manure and activated sludge [22]. According to Edwards [23], the optimum growth of *E. fetida* in different animal and vegetable waste was at temperatures of 25–30°C and at a moisture content range of 75–90%, but these units could vary in different substrates. *E. fetida* is an iteroparous earthworm, with continuous and high reproduction rates, and it should respond to adverse environmental conditions modifying those rates.

The aim of the present work was to know whether biofertilizers such as Azospirillum (Azos), Phosphobacteria (PB), Blue green algae (BGA) and Rhizobium (Rhizo) mixed in the flower waste have any influence on the growth and reproduction of earthworm *E. fetida* population during the vermicomposting process of 80 days.

2. MATERIALS AND METHODS

Healthy adult *E. fetida* were procured from Agricultural research and training centre, Namakkal, Tamil Nadu, India. The worms were brought to the laboratory and reared in plastic containers with partially decomposed organic matter (cow dung).

2.1 Preparation of Vermibeds

Flower waste were collected from gardens, shops, temples etc., were dried under shade for about 10 days and then mixed with dry cow dung at 1:1 ratio (w/w). Plastic circular containers of 28 cm diameter and 30 cm depth with pierced lid for aeration were used for vermicomposting experiment. The flower waste, cow dung mixture was served as bedding as well as food for the earthworms. The bedding substrates was kept for 4 weeks prior to experimentation and watering was done on alternate days for pre-decomposting, microbial degradation and softening of waste. The pre-decomposed substrate was divided into six combination of feed mix

Control (Co) - Substrate alone without Earthworms; A- Substrate with Earthworms; B- Substrate + Earthworms + Azospirillum; C- Substrate + Earthworms + Phosphobacteria (PB); D- Substrate + Earthworms + Blue green algae (BGA); E- Substrate + Earthworms + Rhizobium.

Biofertilizers were mixed at the rate of 1 g per kg of substrate [24]. Triplicates were maintained for all experiment and control. Five weeks old 25 clitellate *E. fetida* were stocked in each experimental container containing 5 kg of substrate material. The moisture level was maintained around 60-80% and throughout the study period by periodic sprinkling of adequate quantity of tap water. To prevent the moisture loss, the experimental containers were covered with jute bags, containers were kept in dark humid place with a temperature of 27 – 30°C. The samples were drawn at a regular interval of 20 days for chemical analysis i.e., Organic carbon, Total nitrogen, phosphorus, potassium and C: N ratio up to 80 days.

The physicochemical parameters of bedding material and vermicompost produced during the experiment were analyzed by using standard methods. The pH was determined using a double distilled water suspension of vermicompost in the ratio of 1:10 (W/V) that was agitated mechanically for 30 min and filtered through whatman no.1 filter paper [25]. Moisture content was estimated after oven drying at 105°C to a constant weight. Total organic carbon (TOC) of the vermicompost was estimated by the method of [26]; Total Kjeldahl nitrogen (TN) by [27]; Available phosphorus by [28], potassium by [29]. C: N ratio was calculated by dividing the percent of carbon with percent of nitrogen.

Vermicomposting Co-efficient (VC) was calculated by using the following formula [30].

$$VC = \frac{\text{Total increase/decrease during vermicomposting (Experiment)}}{\text{Total increase/decrease during composting (Control)}}$$

The growth and reproductive performance of earthworms in different experimental containers were measured at the same interval. The earthworm population was counted by the hand-sorting method as suggested by [20] at 20, 40, 60 and 80 days after the start of the experiment and the material was returned to the same containers.

Growth rate determination, $R = (N_2 - N_1) / T$

Where R = Growth rate; N_1 = Initial earthworm biomass (mg); N_2 = Final earthworm biomass (mg); T = Time period of the experimental day.

Reproductive performance:

This was determined as the Net Reproductive Rate (Nrr) and was computed following [31] as stated below:

$$Nrr = \frac{\text{Total Number of earthworms harvested}}{\text{Total number of earthworm stocked} \times \text{Experimental period (weeks)}}$$

All the Data were analyzed using SPSS statistical package. Differences among different experimental parameters were calculated by using one way ANOVA ($p < 0.05$) and Duncan's multiple range test (DMRT).

3. RESULTS AND DISCUSSION

The vermicomposting co-efficient of physicochemical parameters shows an increased trend in Moisture, Nitrogen, Phosphorus and Potassium, decreases in pH, Temperature, Total Organic Carbon and C: N ratio (Table 1). Maximum pH reduction was recorded in A and minimum in B and C. When compare to temperature a higher reduction in A and least in C and Co, where as highest reduction of TOC observed in E and D, and lowest in Co. A high C: N reduction noted in E and low in A and Co. The increase in earthworms' growth may also be attributed to a low C: N ratio of the pre-decomposed substrate and positive role of bio-inoculants used in the present study [32].

Increased moisture content was recorded in A (82.02%) and B (76.6%) where as reduced level in C (69.58%). The moisture content of the substrate can influence the worm's growth directly or indirectly. The rate of mineralization and decomposition becomes faster with the optimum moisture content [33]. The direct influence concerns the activity (feeding activity) of the worms. According to [34] the feeding activity of certain earthworm species depends on the moisture content of the substrate. *E. fetida* only attains sexual maturity between moisture levels of 65 and 80% [35]. Edwards and Bater [36] reported that optimum moisture content for growth of earthworm's *E. fetida*, *E. eugeniae* and *P. excavatus* was 85% in organic waste management. Hartenstein et al. [12] and Kaplan et al. [17] indicate the greatest biomass and maximum weight gain of earthworms in domestic dung or activated sludge to be achieved at temperatures of 20°C -29°C and at moisture levels of 70%-85%. The major nutrients N (B and E), P (A) and K (B) were increased and decreased in Co. Several studies have shown that when soil N content increases, the abundance, biomass and reproduction of earthworms also increase [37, 38]. The presence of large number of microflora in the gut of earthworm might play an important role in increasing P and K content in the process of vermicomposting. The P content also a direct action of earthworm gut enzymes and indirectly by stimulation of the microflora [39] due to bacterial and faecal

phosphatase activity of earthworms that probably lead towards mineralization and mobilization of phosphorus [40].

Table 1. Vermicomposting coefficient (VC) of different chemical parameters in control and experimental bins

Parameters	Co	A	B	C	D	E
pH	0.9250	0.9189	0.9730	0.9730	0.9459	0.9594
Moisture (%)	1.0000	1.1014	1.0274	0.9337	0.9889	0.9889
Temperature (°C)	1.0000	0.8333	0.875	0.9167	0.8958	0.875
TOC (%)	0.851	0.836	0.826	0.840	0.789	0.776
TN (%)	1.5	1.692	1.709	1.692	1.618	1.709
Phosphorus (%)	2.263	2.765	2.722	2.684	2.594	2.390
Potassium (%)	1.632	1.721	2.969	2.676	2.067	2.232
C:N ratio	1.0000	0.9122	0.8513	0.8466	0.8311	0.7770

Table 2 reveals the earthworm biomass of present study with maximum number in C (439) and E (428) and least in A (207). The increased earthworm biomass with phosphobacteria suggests the dual role of bacteria as food material and in enriching the substrate with phosphorus through phosphorus solubilization. This phenomenon has also been reported by [41]. Various studies have shown that earthworm utilize micro-organisms in their substrates as a food source and can digest them selectively [42, 43]. A highest average body weight was recorded in E (8.8g) and lowest in A (3.3g). Maximum body length was noted in D (1.3cm) and minimum in A and E (0.8cm). The growth rate in earthworms mainly depends upon the microbial populations and availability of nutrients in vermibeds [44]. The presences of fungi during vermicomposting process became additional supplement to the earthworms which contributed to the increased number and weight of the earthworms. Fungi have cell walls composed of chitin that contains high natural protein; amino polysaccharide [45]. In particular, protozoa and fungi are assumed to form a substantial part of their diet [46, 47]. In general, crop residues have different C: N ratios, particle size, proteins and crude fiber content and even some concentration of special plant metabolites i.e., poly-phenols and related substances [48] which may affects the earthworm production rate in the vermibeds.

The earthworms cultured in biofertilizer mixed flower wastes (B,C,D and E) shows high reproductive performance than A in 20th, 40th, 60th and 80th days (Table 3). These results show there was a significant relationship between biofertilizers and earthworm reproductive behaviour. Domínguez et al. [49, 50] found different growth and reproduction rates of *E. andrei* in different diets and they also found that earthworms invested preferentially their energy either to growth or to reproduction depending of the food quality and moreover earthworm's growth is limited by carbon availability [51]. Gunadi et al. [21] found a relationship between increased growth and reproductive rates of *E. fetida* with low C to N ratios of cattle and pig manure, and [32] reported a decrease in growth rates with increased C to N ratio of paper mulch. A better worm growth pattern in *E. fetida* was due to a good supply of easily available metabolizable organic matter, non assimilated carbohydrates and even low concentration of growth retarding substances in vermibeds. The survival of earthworms in waste decomposing sub system mainly depends on physical and initial chemical profile of the feed stuff [52, 53]. The survival, biomass production and reproduction of earthworms are the best indicator to evaluate the vermicomposting process [30]. The earth worm biomass has been increased 7.28 to 16.56 times in the biofertilizers enriched flower waste vermicomposting process of 80 days. Elvira et al. [4] reported earthworm increased 2.2 to 3.9 times total biomass in the combination of paper mill sludge with cattle wastes.

Table 2. Relative growth rate of *Eisenia fetida* in biofertilizers enriched flower waste vermicompost (mean±S.D)

Experiments	Number of Worms		Average Body Weight (g)		Average Body Length(cm)	
	Initial	Final	Initial	Final	Initial	Final
A	25±0.00	206.6±1.140 ^d	8.74±0.051 ^a	12.48±1.134 ^b	6.74±1.920 ^a	6.78±0.083 ^c
B	25±0.00	336.4±2.073 ^b	8.554±0.0357 ^a	15.5±0.20 ^a	5.82±0.148 ^a	7.96±0.712 ^a
C	25±0.00	436.4±1.673 ^a	6.694±0.450 ^b	13.40±2.140 ^b	6.20±0.187 ^a	7.24±0.167 ^b
D	25±0.00	280.2±0.320 ^c	6.14±0.320 ^c	11.7±0.412 ^b	6.42±1.132 ^a	7.38±0.402 ^b
E	25±0.00	431.2±4.868 ^a	6.28±0.148 ^c	16.0±1.581 ^a	6.42±0.506 ^a	7.38±0.192 ^b

Table 3. Reproductive performance of *Eisenia fetida* during vermicomposting (80 days)

Experiments	Initial stocking	20 th day		40 th day		60 th day		80 th day	
		EW	RP	EW	RP	EW	RP	EW	RP
A	25.0±0.00	35.6±1.516 ^d	0.47±0.0158 ^d	82.6±1.516 ^e	0.584±0.427 ^d	151.6±2.701 ^e	0.72±0.109 ^d	205.8±1.303 ^e	0.66±0.015 ^d
B	25.0±0.00	74.6±2.966 ^b	1.058±0.019 ^b	152.4±1.673 ^c	1.074±0.015 ^b	183.4±2.073 ^c	0.84±0.01 ^c	334.5±1.581 ^c	0.954±0.036 ^b
C	25.0±0.00	82.8±1.920 ^a	1.19±0.514 ^a	183.8±2.588 ^a	1.28±0.050 ^a	252.8±1.788 ^b	1.16±0.0244 ^b	435.6±2.509 ^a	1.364±0.019 ^a
D	25.0±0.00	54.6±2.073 ^c	0.754±0.078 ^c	131.6±0.894 ^d	0.916±0.078 ^c	175.8±1.303 ^d	0.844±0.033 ^c	278.6±4.929 ^d	0.874±0.011 ^c
E	25.0±0.00	56.0±1.581 ^c	0.758±0.049 ^c	174.0±2.000 ^b	1.226±0.011 ^a	292.6±3.209 ^a	1.356±0.025 ^a	427.6±4.505 ^b	1.388±0.021 ^a

4. CONCLUSION

In the present study highest earthworm biomass were observed in C, E and least in A, biomass has been increased 7.28 to 16.56 times in the biofertilizers enriched flower waste vermicomposting process of 80 days. Based on the present study we conclude biofertilizers enriched flower waste vermicomposting shows better results of earthworm biomass and reproductive performance influence by the biofertilizers added in the substrate material. There are opportunities to study the influence of biofertilizers and substrate materials inter relating with environmental factors in the field of vermiculture. Further work is required to establish the exact biofertilizer for improve the production of earthworms for sustainable vermicomposting / vermiculture operations.

ACKNOWLEDGEMENT

The authors would like to gratefully thank Dr. G. Manimekalai, Head, Department of Zoology, Kandasamy Kandar College, P.Velur, Tamil Nadu for permit to conduct this study, authors thank the Administrative authorities of Kandasamy Kandar College for providing infrastructure facilities.

COMPETING INTERESTS

Authors have declared that no competing interests exist

REFERENCES

1. Norbu T. Pretreatment of municipal solid waste by windrow composting and vermicomposting. A thesis for the degree of Master of Science, August 2002. Asian Institute of Technology, Thailand. 2002;1-103.
2. Nagavallema KP, Wani SP, Stephane L. Vermicomposting: recycling wastes into valuable organic fertilizer". Journal of SAT Agricultural Research. 2006;2(1):1-17.
3. Benitez E, Nogales R, Elvira C, Masciandaro G, Ceccanti B. Enzyme activities as indicators of the stabilization of sewage sludge composting with *Eisenia foetida*. Bioresource Technology. 1999;67:297-303.
4. Elvira C, Sampedro L, Benitez E, Nogales R. Vermicomposting of sludges from paper mill and dairy industries with *Eisenia andrei*: A pilot scale study. Bioresource Technology. 1997;63:205-211.
5. Kumar DS, Jagadeesan L. Comparative decomposing efficacy of three earthworm species on paper mill solid waste and macronutrient analysis. Environment and Ecology. 2010;28(1):212-215.
6. Gratelly P, Benitez E, Elvira C, Polo A, Nogales R. Stabilization of sludges from a dairy processing plant using vermicomposting. In: Rodriguez-Barrueco C (ed) Fertilizers and environment. Kluwer, Netherlands. 1996;341-343.
7. Nogales R, Celia C, Benitez E. Vermicomposting of winery wastes: a laboratory study. Journal of Environment Science Health Part B. 2005;40:659-673.
8. Maboeta MS, Van Rensburg L. Vermicomposting of industrially produced woodchips and sewage sludge utilizing *Eisenia fetida*. Ecotoxicology and Environmental Safety. 2003;56:265-270.

9. Kumar DS, Anbuganapathy G. Effect of Biofertilizer application on flower waste subjected to vermicomposting using *Eisenia fetida* and nutrient analysis of vermicompost. *Scientific Transaction in Environment and Technovation*. 2010;3(3):134-138.
10. Kaushik P, Garg VK. Dynamics of biological and chemical parameters during vermicomposting of solid textile mill sludge mixed with dung and agricultural residues. *Bioresource Technology*. 2004;94:203–209.
11. Garg VK, Kaushik P. Vermistabilization of textile mill sludge spiked with poultry droppings by an epigeic earthworm *Eisenia fetida*. *Bioresource Technology*. 2005;96:1063-1071.
12. Kaplan DL, Harstenstein R, Neuhauser EF, Maleckii MR. Physico-chemical requirements in the environment of the earthworm *Eisenia fetida*. *Soil Biology and Biochemistry*. 1980;12:347-352.
13. Schulz E, Graft O. Zur Bewertung von Regenwurmmehl aus *Eisenia fetida* (Savigny 1926) als Eiweissfuttermittel. *Landbauforsch Volkenrode*. 1977;27:216-218.
14. Venter JM, Reinecke AJ. The life-cycle of the compost worm *Eisenia fetida* (Oligochaeta). *South African Journal of Zoology*. 1988;23:161-165.
15. Reinecke AJ, Venter JM. Moisture preferences, growth and reproduction of the compost worm *Eisenia foetida* (Oligochaeta). *Biology and Fertility of Soils*. 1987;3:135-141.
16. Reinecke AJ, Kriel JR. Influence of temperature on the reproduction of the earthworm *Eisenia fetida* (Oligochaeta). *South African Journal of Zoology*. 1981; 16: 96-100.
17. Hartenstein R, Neuhauser EF, Kaplan DL. Reproductive potential of the earthworm *Eisenia foetida*. *Oecologia*. 1979;43:329-340.
18. Tomlin AD, Miller JJ. Development and fecundity of the manure worm, *Eisenia foetida* (Annelida: Lumbricidae), under laboratory conditions. In: Dindal, D.L. (ed) *Soil Biology as Related to Land Use Practices*. Office of Pesticides and Toxic Substances, EPA, Washington, D.C. 1980;673–678.
19. Venter JM, Reinecke AJ. Can the commercial earthworm *Eisenia fetida* (Oligochaeta) reproduce parthenogenetically. *Rev Ecol Biol Sol*. 1987;24:157-170.
20. Lee KE. *Earthworms - Their ecology and relationships with soils and land use*. Academic Press, Sydney; 1985.
21. Gunadi B, Edwards CA, Blount C. The influence of different moisture levels on the growth, fecundity and survival of *Eisenia foetida* (Savigny) in cattle and pig manure solids. *Euro J Soil Biol*. 2003;39:19–24.
22. Loehr RC, Neuhauser EF, Malecki MR. Factors affecting the vermistabilization process. *Water Research*. 1985;19(10):1311–1317.
23. Edwards CA. The use of earthworms in the breakdown and management of organic wastes. – In: Edwards, C.A. (ed.): *Earthworm Ecology*. St. Lucie Press, Boca Raton. 1998:327-351.
24. Subramaniam P. *Vermicomposting Technology*, Chapter.3. Perspectives of Organic Agriculture. Published by the Director, CAS Department of Agronomy, Tamil Nadu Agriculture University, Coimbatore; 2006:126- 133.
25. McLean EO. Soil pH and lime requirement. *Methods of Soil Analysis. Chemical and Microbiological Properties*. 2nd (Eds), Number 9 (Part 2) in series. American Society of Agronomy, Madison. 1982:199-224.
26. Nelson DW, Sommers LE. Total carbon, organic carbon and organic matter. In: *Methods of soil analysis. Part 2. Chemical and microbiological properties*. A.L. Page, R.H; 1982.

27. Bremner JM, Mulvaney CS. Nitrogen-Total In: Methods of Soil Analysis (A. L. Page et al., Ed.), Agronomy Monograph 9, Part II, 2nd ed. American Society of Agronomy, Madison. 1982;595-624.
28. Anderson JM, Ingram JSI (Eds.) Tropical Soil Biology and Fertility: A Handbook of Methods. CAB International, UK. 1993:95-97.
29. Simard RR. Ammonium acetate extractable elements. In: Martin, R., Carter, S. (Ed), Soil Sampling and Methods of Analysis. Lewis Publisher, FL, USA. 1993:39-43.
30. Suthar S. Potential utilization of guar gum industrial waste in vermicompost production. Bioresource Technology. 2006;97:2474-2477.
31. Dynes RA. Earthworms-Technology information to enable the development of earthworm production. Rural Industrial Research Development Corporation Publication No. 03/085. 2003:1-3.
32. Ndegwa PM, Thompson SA. Effects of C-to-N ratio on vermicomposting of biosolids. Bioresource Technology. 2000;75:7-12.
33. Singh NB, Khare AK, Bhargava DS, Bhattacharya S. Optimum moisture requirement during vermicomposting using *Perionyx excavates*. Applied Ecology and Environmental Research. 2004;2:53-62.
34. Mitchell MJ, Mulligan RM, Hartenstein R, Neuhauser EF. Conversion of sludges into "top- soils" by earthworms. Compost Science. 1977;18:28-32.
35. Reinecke AJ, Venter JM. The influence of moisture on the growth and reproduction of the compost worm *Eisenia fetida* (Oligochaeta). Recue d' Ecologie et de Biologie du Sol. 1985;22:473-481.
36. Edwards CA, Bater JE. The use of earthworms in environmental management. Soil Biology and Biochemistry. 1992;24(12):1683-1689.
37. Jimenez JJ, Moreno AG, Decaens T, Lavelle P, Fisher MJ, Thomas RJ. Earthworm communities in native savanna and man-made pastures of the Eastern Plain of Colombia. Biology and Fertility of Soils. 1998;28:101-110.
38. Bohlen PJ, Parmelee RW, Allen MF, Kettering QM. Differential effects of earthworms on nitrogen cycling from various nitrogen-15-labelled substrates. Soil Science Society of America Journal. 1999;63:882-890.
39. Satchell JE, Martein K. Phosphate activity in earthworm faeces. Soil Biology Biochemistry. 1984;16:191-194.
40. Edwards CA, Lofty JR. Biology of Earthworms. Chapman and Hall, London; 1972.
41. Kumar V, Narula N. Solubilization of inorganic phosphates and growth emergence of wheat. Biology and Fertility of Soils. 1999;28:301-305.
42. Curry JP, Schmidt O. A feeding ecology of earthworms. A review. Pedobiologia. 2006;50(6):463-477.
43. Subler S, Kirsch AS. Spring dynamics of soil carbon, nitrogen and microbial activity in earthworm hidden in a no till cornfield. Biology and Fertility of Soils. 1998;26(3):243-249.
44. Suthar S. Reproduction and growth of earthworms in cattle waste solid. In: Dahnof, L.T (Ed), Animal Reproduction: New Research Developments, Nova Science Publishers, Inc., USA; 2009:284-294.
45. Kumar. Chitin and chitosan fibres: A review. Bulletin of Material Science. 1995;22(5):905-915.
46. Brown GG. How do earthworms affect microfloral and faunal community diversity? Plant and Soil Science. 1995;170:209-231.
47. Bonkowski M, Schaefer M. Interactions between earthworms and soil protozoa: a trophic component in the soil food web. Soil Biology and Biochemistry. 1997;29:499-502.

48. Ganesan PS, Gajalakshmi S, Abbasi SA. Vermicomposting of leaf litter of acacia (*Acacia auriculiformis*) possible roles of reactor geometry, polyphenols and lignin. *Bioresource Technology*. 2009;100(5):1819-1827.
49. Domínguez J, Briones MJL, Mato S. Effect of the diet on growth and reproduction of *Eisenia andrei* (Oligochaeta, Lumbricidae), *Pedobiologia*. 1997;41:566–577.
50. Domínguez J, Edwards CA, Webster M. Vermicomposting of sewage sludge: effect of bulking materials on the growth and reproduction of the earthworm *Eisenia andrei*, *Pedobiologia*. 2000;44:24–32.
51. Tiunov AV, Scheu S. Carbon availability control the growth of detritivores (Lumbricidae) and their effect on nitrogen mineralization, *Oecologia*. 2004;138:83–90.
52. Butt KR. Utilization of solid paper mill sludge and spent brewery yeast as a feed for soil dwelling earthworms. *Bioresource Technology*. 1993;44(2):105-7.
53. Suthar S. Pilot scale vermireactors for sewage sludge stabilization and metal remediation process: comparison with small scale vermi-reactors. *Ecological Engineering*. DOI: 10. 1016/j. ecol eng. 2009.12.016.

© 2013 Kumar et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<http://www.sciencedomain.org/review-history.php?iid=239&id=9&aid=1574>